



Improving outcomes through Collaboration in Osteosarcoma

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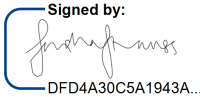



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Please note: This trial protocol must not be applied to patients outside the ICONIC study. Cancer Research UK & UCL Cancer Trials Centre (UCL CTC) can only ensure that approved study investigators are provided with amendments to the protocol.

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ABBREVIATIONS

ACBS	Aarhus composite biomarker score
AE	Adverse Event
ALP	Alkaline phosphatase
ALT	Alanine transaminase
AP	Doxorubicin/Cisplatin chemotherapy
AR	Adverse Reaction
ASA classification	American Society of Anesthesiologists classification of fitness for surgery system classification
AST	Aspartate aminotransferase
BCRT	The Bone Cancer Research Trust
BRCA	BReast CAncer susceptibility gene
ccfDNA	Circulating cell free DNA
cfDNA	Cell free DNA
CI	Chief Investigator
CITA	CRUK Accelerator grant on Cancer ImmunoTherapy
CM	Centralised monitoring
CN	Copy number analysis
CNA	Copy Number Alterations
CNV	Copy number variations
CPI	Checkpoint inhibitor
CRP	C-reactive protein
CR UK	Cancer Research UK
CT	Computerised Tomography
CTC DNA	Circulating tumour cell DNA
CTCAE	Common Terminology Criteria for Adverse Events
ctDNA	Circulating tumour DNA
DARWIN 1	Deciphering Afatinib Response and Resistance with INtratour heterogeneity study
DEPArray™	DEPArray enables manipulation and recovery of rare cells by combining image-based cell selection with DEP movement single cell sorter
ECMC GCLP	Experimental Cancer Medicine Good Clinical Laboratory Practice Facility
eCRF	Electronic Case Report Form
EDTA	Ethylene Diamine Tetra Acetate
EORTC QLQ-C30	European Organization for Research and Treatment of Cancer Quality of Life Questionnaire-Core 30
EURAMOS-1	A randomized trial of the European and American Osteosarcoma Study Group to optimize treatment strategies for resectable osteosarcoma based on histological response to pre-operative chemotherapy
FFPE	Formalin fixed paraffin embedded blocks
FGFR1	Fibroblast Growth Factor Receptor 1
fMRI	Functional MRI
FOSTER	Fighting OsteoSarcoma Through European Research
GCP	Good clinical practice
GDPR	EU General Data Protection Regulation 2016 (EU)2016/679
GeCIP	Genomics England Clinical Interpretation Partnership
GRC	Global rating of change Scale

Hb	Haemoglobin
HRA	Health Research Authority
HRD	Homologous Recombination Deficiency
ICH GCP	International Conference of Harmonisation-Good Clinical Practice
ICONIC	Improving outcome through Collaboration in Osteosarcoma
ICGC	International Cancer Genome Consortium
IGF(R)	Insulin-like Growth Factor (receptor)
IO	Immune Oncology
IP	Intellectual Property
ISAP	Independent Scientific Advisory Panel
ISF	Investigator Site File
ISRCTN	International Standard Randomised Controlled Trial Number
IV	Intravenous
LDH	Lactate dehydrogenase
LN	Lymph node
MAMS	Multiarm, multistage study
MAP	High dose methotrexate, doxorubicin, cisplatin
MDM	Medical decision making
MDT	Multi Disciplinary Team
MT1-MMP	Membrane type 1 metalloprotease enzyme contributing to angiogenesis.
mNCA	model Non-Commercial Agreement
MRI	Magnetic Resonance Image
NCITA	National Cancer Imaging Translational Accelerator
NCRI	National Cancer Research Institute
NICE	National institute for health and care excellence
NIHR	National institute for health research
Non CTIMP	Clinical Trial not involving an Investigational Medicinal Product
ORC	Osteosarcoma Research Consortium
OS	Osteosarcoma
PAs	Patient Advocates
PBMCs	Peripheral Blood Mononuclear Cells
PET/CT	Positron Emission Tomography - Computed Tomography scan
PI	Principal Investigator
PIS	Patient information sheet
PPIE	Patient and Public Involvement and Engagement
PRO	Patient Reported Outcome
QoL	Quality of life
R&D	Research & development
rEECur	An international randomised controlled trial of chemotherapy for the treatment of recurrent and primary refractory Ewing sarcoma.
REC	Research Ethics Committee
RNOH	Royal National Orthopaedic Hospital
RT	Radiotherapy
RTD	Routes to Diagnosis
SAR	Serious Adverse Reaction
SAM	Sarcoma Assessment Measure
SOC	Standard of Care
TAA	Tumour associated antigens

TESS	Toronto Extremity Salvage Score of function
TMF	Trial Master File
TMG	Trial Management Group
TRACERx	TRACKing non-small cell lung Cancer Evolution through therapy [Rx]
TSC	Trial Steering Committee
TYA	Teenage & young adult cancers
UCL	University College London
UCL CTC	CR UK and UCL Cancer Trials Centre
UCLH	University College London Hospital
WB MRI	Whole body MRI
WGS	Whole genome sequencing
WTSI	Wellcome Trust Sanger Institute

1. PROTOCOL SUMMARY

1.1. Summary of Study Design

Title:	Improving outcomes through Collaboration in Osteosarcoma
Short Title/acronym:	ICONIC
Sponsor name & reference:	University College London Reference: 18/0248
Funder name & reference:	Bone Cancer Research Trust
Clinicaltrials.gov no:	NCT04132895
Design:	Prospective observational cohort study.
Overall aim:	To improve treatment and outcomes for patients with osteosarcoma (OS) by establishing a clinically annotated cohort of newly diagnosed patients with OS with longitudinal collection of biospecimens.
Primary objective:	To obtain high quality clinical data and biospecimen collection from OS patients to develop potential new osteosarcoma trials.
Secondary objectives:	<p>To enrol 300-350 patients to address the following questions:</p> <p>CLINICAL OBJECTIVES</p> <ul style="list-style-type: none"> Describe the impact of variation in management of patients with OS across treatment sites in the UK Assess whether tumour margin and response to chemotherapy predict local recurrence in OS <p>BIOLOGICAL OBJECTIVES</p> <ul style="list-style-type: none"> Assess whether OS tumour and immune models grown from OS patients can be used to identify new therapeutic targets Explore whether new candidate immunotherapy drug targets for OS can be discovered based on cellular and genomic analysis Collect peripheral blood mononuclear cells (PBMCs) from OS patients to inform immune studies Develop patient derived explants, primary cell cultures, cell lines and organoid models from fresh/frozen patient tumour tissue to investigate OS biology and validate new candidate drug targets Use formalin-fixed paraffin-embedded (FFPE) samples to evaluate genetic and immune biomarkers and aid development of novel therapeutic targets and trials Create a legacy resource of OS immune oncology (IO) data and patient models to accelerate future collaborative research efforts

	<p>CIRCULATING BIOMARKERS</p> <ul style="list-style-type: none"> • Assess whether circulating biomarkers can be used as prognostic markers and to predict burden of disease, response to therapy and outcome • Determine the prevalence of Germline DNA alterations in UK OS patients <p>IMAGING OBJECTIVES</p> <ul style="list-style-type: none"> • Determine if standard of care imaging can be used to conduct imaging biomarker studies through establishment of an imaging repository • Examine the effects of tumour heterogeneity and clonal evolution on chemotherapy response and patient outcome and determine if this can be used to stratify patients for therapy <p>PATIENT EXPERIENCE</p> <ul style="list-style-type: none"> • Assess the value of the Sarcoma Assessment Measure (SAM) for longitudinal assessment of patient quality of life and identify predictors of variance in patient reported outcomes (PROs) • Describe the routes and timescales of diagnostic pathways for patients with OS, and assess whether this has any impact on patient outcome
<p>Target accrual:</p>	<p>A minimum of 350 patients (including those already recruited in ICONIC).</p>
<p>Eligibility criteria:</p>	<p><i>Inclusion criteria</i></p> <ul style="list-style-type: none"> • New histological diagnosis of osteosarcoma or in the absence of osteoid seen on biopsy, pathology and imaging supportive of a diagnosis of osteosarcoma • Written informed consent of patient and/or parent/legal guardian or written confirmation of opinion of person with duty of care where applicable <p><i>Exclusion criteria</i></p> <ul style="list-style-type: none"> • Diagnosis more than four months prior to registration
<p>Number of sites:</p>	<p>At least 25 sites representing OS specialist oncology and surgical sites across the UK</p>
<p>Treatment summary:</p>	<ul style="list-style-type: none"> • Treatment will be given as per usual standard of care. • No treatments are specified by this protocol.

<p>Patient study activities:</p>	<p>Clinical data collection</p> <p>Clinical data will be collected on a regular basis. Data to be collected include baseline characteristics, treatment details, results of key investigations (including blood tests and scans), patient status (including performance status) and outcome.</p> <p>Tissue and blood collection</p> <ul style="list-style-type: none"> • Tissue for FFPE and fresh freezing according to local policy will be collected during routine procedures at diagnosis, at surgery and at relapse if clinically indicated. • Optional study specific biopsies will be requested at relapse in cases where a biopsy is not clinically indicated. • Blood samples will be collected from patients receiving chemotherapy and/or surgery at the same timepoints as tissue, with an additional sample presurgery and further optional samples. <p>Patient Reported Outcomes (PROs)</p> <p>Sarcoma Assessment Measure (SAM) questionnaire</p> <p>Patients ≥ 13 years of age will be requested to complete a Sarcoma Assessment Measure (SAM) questionnaire along with TESS, EORTC-QLQ-C30 and Global rating of change (GRC) at registration, annually and at relapse.</p> <p>Prediagnostic data collection</p> <ul style="list-style-type: none"> • Patients ≥ 13 years will be requested to complete a questionnaire at registration detailing symptoms and routes to diagnosis. • For patients < 13 years, the parents/legal guardian will be requested to complete the questionnaire at registration detailing symptoms and routes to diagnosis.
<p>Central laboratory analyses:</p>	<p>Tissue</p> <p>Frozen tumour tissue will be analysed as part of the 100,000 Genomes project, or through NHS England where possible for WGS, further analyses may be undertaken including RNA and epigenetic analysis as part of Osteosarcoma Research Consortium (ORC).</p> <p>Fresh/frozen tumour tissue will be used to establish tumour explant, primary cell cultures, cell lines and patient derived organoid models, for validation of new candidate therapeutic targets, including co-culture experiments of organoids/explants together with matched autologous immune cell samples and compound screenings.</p>

	<p>FFPE tissue will be used to:</p> <ul style="list-style-type: none"> • Determine feasibility of DNA/RNA extraction for validation studies • Validate findings from WGS and other analyses including molecular targeted sequencing and copy number analysis • Conduct spatial transcriptomics analysis and high dimensional immunohistochemistry analysis <p>Blood</p> <p>Blood samples will be collected to:</p> <ul style="list-style-type: none"> • Investigate germline alterations • Evaluate two circulating tumour cell (CTC) isolation and characterisation methods: <ul style="list-style-type: none"> • Parsortix™ and DEPArray™ • Flow cytometry using MT1-MMP Ab • Store for potential methylation profiling, genetic analysis of circulating tumour DNA (ctDNA) and future research <p>To Inform immune studies blood samples will also undergo Ficoll based centrifugation to obtain fresh PBMCs.</p>
Duration of recruitment:	To 31 January 2025
Duration of follow up:	Patients will be followed up for minimum of two years, with a median of approximately 56 months follow up.
Definition of end of study:	The end of study will be declared two years after recruitment of the last patient.

1.2. Study Summary

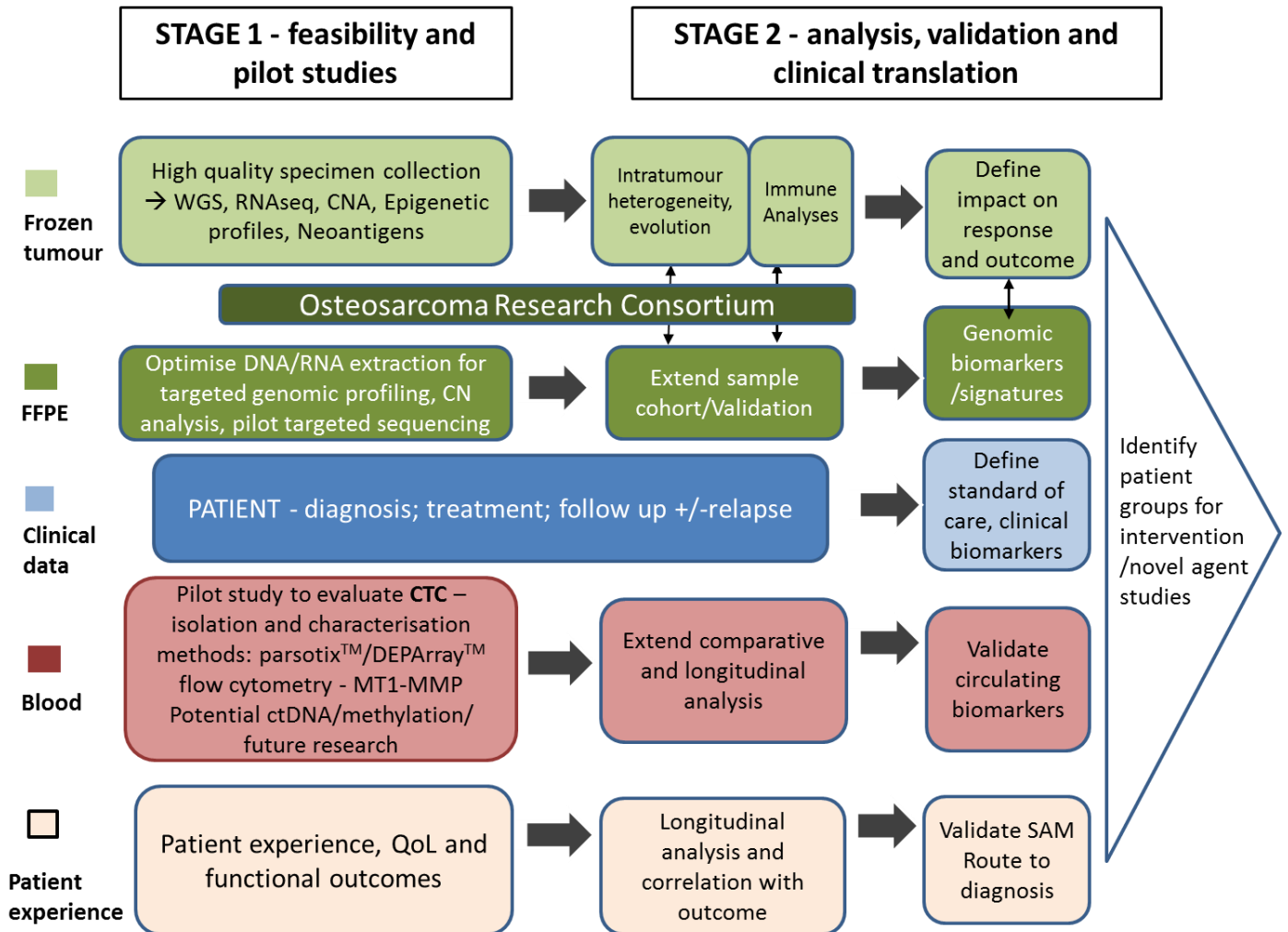
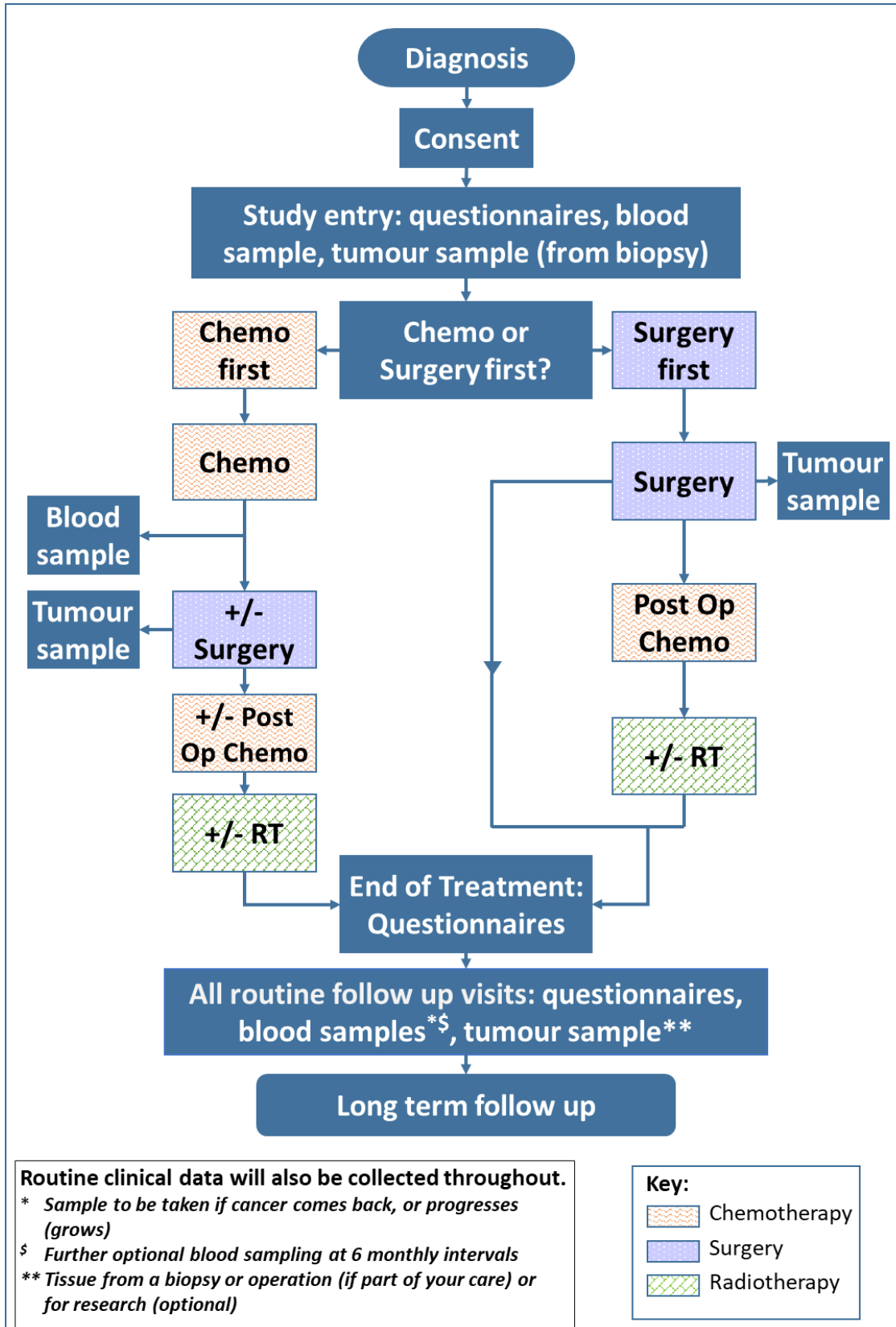


Figure 1: Summary Schema of ICONIC and output

- FFPE = formalin-fixed paraffin-embedded
- WGS = whole genome sequencing
- CNA = copy number alterations
- CTC = circulating tumour cell
- PRO = patient reported outcome
- SAM = sarcoma assessment measure

Study flowchart



Sample Schedules

SCHEDULE: PATIENTS RECEIVING SURGERY ALONE +/- ADJUVANT CHEMOTHERAPY	Within 28 days after registration	Surgery	End of Treatment	Follow up	Relapse (local recurrence or metastases)
Whole blood sample for germline DNA	X				
Whole blood sample for ctDNA (optional for all patients)	X		X	X ¹	X
FFPE tumour tissue	X	X			X ²
Provide information on frozen tumour tissue ³	X	X	X		X ²
Frozen tissue samples (send on request)	X	X			X

1. To be collected at clinic visit during follow up no more than 6 monthly.
2. Collect if patient consents to this optional biopsy (if not already collected as part of SOC)
3. Where sample stored, reason sample not taken if applicable

SCHEDULE: PATIENTS RECEIVING NEOADJUVANT CHEMOTHERAPY	Baseline: within 28 days after registration	During Treatment		End of Treatment	Follow up	Relapse (local recurrence or metastases)
		End of Neoadjuvant chemotherapy	Surgery			
Whole blood sample for germline DNA	X					
Whole blood sample for CTCs	X	X	(X) ¹			X
Whole blood sample for ctDNA (optional for all patients)	X	X	(X) ¹	X	X ²	X
PBMCs (UCLH only)	X	X	(X) ¹	X		X
FFPE tumour tissue	X		X			X ³
Provide information on frozen tumour tissue ⁴	X		X	X		X ³
Frozen tissue samples (send on request)	X		X			X ³

1. Can collect before primary resection surgery if not taken at end of neoadjuvant treatment
2. To be collected at clinic visit during follow up no more than 6 monthly.
3. Collect if patient consents to this optional biopsy (if not already collected as part of SOC).
4. Where sample stored, reason sample not taken if applicable

2. BACKGROUND AND RATIONALE

Osteosarcoma (OS) is the most common bone sarcoma, accounting for approximately 30% of all bone sarcoma diagnoses. In England, it accounts for approximately 130 cases per year across all ages (Table 1). Age-specific overall survival incidence rates are generally bimodal, with the first peak in adolescence and a second peak in patients over 60 years of age. Treatment is multimodal, including multi-agent chemotherapy and surgery to the primary site and to metastases where appropriate. Outcome, however, has improved little over the past 25 years with 5-year survival rates remaining about 42% for all ages and stages in the UK. Five-year relative survival rates are significantly higher for patients <40 years of age (52%) compared with patients over 40 years (25%)¹. Survival is also dependent on the primary site of disease, and is poor for patients with non-extremity tumours, at 36% for patients under the age of 40 years and as low as 6% for patients over 40 years. Outcome for patients with metastatic disease at diagnosis remains equally poor²⁻⁵. The use of chemotherapy and surgery varies according to factors such as age, primary site, stage and performance status with approximately two thirds currently receiving these treatments in England each year (Table 1).

Age (years)	< 40	%	40-59	%	60 +	%	Total No.	%
Total	86	63	20	15	31	22	136	100
Chemotherapy	70	82	13	65	6	19	89	66
Surgery	64	75	13	65	13	42	90	66

Table 1. Average number of patients diagnosed per year with OS receiving chemotherapy and/or surgery in England (2011-2015)[§]

[§](National Cancer Registration and Analysis Service, personal communication)

The most recent randomised trial in osteosarcoma, EURAMOS-1 (NCT00134030), was an excellent example of international collaboration to seek improved treatments for a rare cancer. Despite the participation of over 2000 patients, no changes to the standard of care for osteosarcoma resulted^{6,7}. Thus, there continues to be an unmet need to identify new approaches, including biomarkers and novel, innovative and targeted therapeutic approaches if outcomes are to improve. This includes seeking improved systemic treatments and better approaches to management of the primary tumour. Failure of local control (local recurrence) is associated with high levels of morbidity and poor survival⁸. New imaging modalities such as CT-PET, MRI-PET and functional MRI (fMRI) could be useful to guide resection or detect local recurrence but there is insufficient evidence to identify a clear role for use in routine clinical practice⁹. Further prospective study of newly described prognostic systems and identification of other biomarkers of local failure are needed to assist in decision making about morbid surgery and to determine the role of technical advances in radiation oncology which may be of benefit to patients with OS¹⁰.

OS clinical trials to date have focussed on a limited number of outcomes, usually in younger patients with localised extremity disease. Thus, addressing complex inter-related questions has not been possible and several sub-populations have been excluded from study so limiting opportunities to improve the standard of care. These include: those with widely metastatic disease; less common anatomic locations such as pelvis, spine and craniofacial bones; those arising on a background of skeletal dysplasia or underlying genetic predisposition; and finally, the 50% of OS arising in patients over 40 years. Little is currently known about factors influencing treatment decisions in this population and how consistently a standard of care is applied. The effect of treatment on quality of life (QOL), patient reported outcome and other

performance indicators is also not well described or understood. Overall, there is a need to broaden the ambition and scope of OS research while improving access for all patients.

2.1. Osteosarcoma biology and translational research

Therapeutic development in OS has been hindered by the heterogeneous nature of the disease and limited understanding of the molecular and cellular mechanisms that influence its pathogenesis and survival. Currently no molecular sub classification of OS exists and stratification of therapy based on genetic or molecular biomarkers is not possible. Research is under way to identify new biological insights but greater coordination and better linkage to clinical application will optimise the return and improve the delivery timelines of these efforts. In particular, access to systematically collected tissue linked to comprehensive clinical data for all OS is a pressing need to establish a resource of clinically relevant *in vitro* models (primary cultures, cell lines, organoids, explants) that can be used for discovery and validation of new candidate therapeutic targets. Access to these models will foster translational research to improve our understanding of mechanisms of resistance to chemotherapy and susceptibility to immune-based approaches. A more comprehensive characterisation of the transcriptomic and proteomics profiles of patient samples correlated to clinical data will open up new avenues for novel patient stratification strategies.

2.2. Identification of genomic targets

WGS and exome analysis is beginning to provide better insights into the genetic complexity and heterogeneity of OS. One such study, based on whole exome sequencing revealed a **BRCA-ness phenotype** that if validated and demonstrated to be functional could be used to identify patients for targeted therapy¹¹. More recent data using whole genome analysis has however refuted this claim, with only one of 37 patients having an “HRD detect” phenotype, developed by the Sanger Institute as a measure of “BRCAness” thus further information is required to inform development of clinical trials¹². The largest WGS project to date, from the International Cancer Genome Consortium (ICGC), has identified distinct patterns of genomic rearrangement in OS but also recurrent mutation of **Insulin-like Growth Factor** (IGF) signalling genes, present in approximately 14% of patients¹³. IGFR inhibition has previously been investigated in sarcoma with a small proportion of patients benefitting, but no biomarker identifies those most likely to benefit and no studies are currently ongoing. **Fibroblast Growth Factor Receptor 1** (FGFR1) amplification appears to be related to chemoresistance in OS: 20% of patients with a poor histological response harbour the amplification¹⁴. These findings support further investigation in OS of these targets but prospective validation in diagnostic samples and correlation with outcome is now necessary to determine a role in stratification or therapy more clearly.

2.2.1. Intratumour heterogeneity and tumour evolution

Intratumour genetic heterogeneity, where different cells in the same tumour show distinct genomic profiles, is increasingly recognised as a cause of treatment failure, as it allows tumours to adapt and evolve to escape therapy¹⁵. A Cancer Research UK (CR UK) funded study in patients with primary lung cancer, TRACERx (TRACKing non-small cell lung Cancer Evolution through therapy [Rx] (NCT01888601)) is studying the genomic landscape of lung cancer and how it changes through the disease course by taking multiple tumour and blood samples from patients at intervals from diagnosis to relapse¹⁶. This study has already significantly contributed to our understanding of cancer evolution, demonstrating that intratumour heterogeneity mediated through chromosome instability is associated with an increased risk of recurrence or death¹⁶. The study is also investigating the effects of heterogeneity on response to

platinum chemotherapy and, more recently, enabled investigation into tumour interaction with the immune system with well publicised new discoveries that will support precision medicine¹⁷. This model is expected to be applicable to other cancer types, including OS, but requires significant investment.

Professor Flanagan is leading a project '**The Osteosarcoma Research Consortium (ORC)**' within UCL, funded by a generous donation from the Tom Prince Research Trust. This study is investigating multiple aspects of OS including genetic and epigenetic heterogeneity over space and time and has commenced recruitment of patients from the London Sarcoma Service, The Robert Jones Agnes Hunt NHS Trust in Oswestry, and Ireland, and has aligned this study with the 100,000 Genomes project and ongoing sequencing through NHS England. The study includes WGS being performed on high quality patient samples including diagnostic biopsies, resection specimens (for multiple region WGS), and metastatic and relapse samples. Copy number alterations, RNA sequencing and processing of samples for epigenetic studies will be performed on the same samples in the Flanagan laboratory. Some tumour samples are also being prepared for single cell genomic analysis as a pilot project. The multi-'omic' analysis of the data will be undertaken by the research consortium comprising members of the Genomics England Clinical Interpretation Partnership (GeCIP) which at present includes Professor Campbell and Dr Behjati – Wellcome Trust Sanger Institute (WTSI), Professor Stephan Beck and Dr Pillay – UCL Cancer Institute, Dr Van Loo – Francis Crick Institute, and clinicians involved in care of the patients. Data analysis of the osteosarcomas will be performed alongside that of the other 400-500 sarcomas submitted to the Genomics England 100,000 Genomes Project, the data of which became available in late 2018 with analysis expected to start in earnest in 2019. To date samples from 90 OS patients have been submitted for WGS and it is estimated that a further 50 patients will be recruited as part of this initiative which will provide the first detailed insights into OS heterogeneity and tumour evolution by 2020.

As part of the ORC, the immune landscape of OS is also being studied and specifically whether the genomic structural variation in OS is a source of neoantigens and therefore potential therapeutic targets. Associations will be sought between both the number and identity of neoantigens and phenotypic data available as well as their association with RNA expression profiles and host immune system response. This project will benefit from the CRUK Accelerator grant on Cancer ImmunoTherapy (CITA) in UCL working with Prof Herrero (Bill Lyons Informatics Centre) and Professor Quezada, a world leading authority on immune checkpoint biology in cancer. This work is particularly relevant in view of disappointing initial results from checkpoint inhibitor trials in OS and will facilitate identification of patients for appropriate immune-based therapies.

Not all patients however have sufficient high quality frozen tissue for WGS, particularly from small diagnostic biopsies, and the above project is currently limited to patients recruited to the 100,000 Genomes project. Consistent coordinated collection of clinically annotated tumour samples within ICONIC will facilitate application for funding for analysis of additional samples and for validation studies on the basis of preliminary data. For patients without adequate frozen tumour samples, the potential to use formalin-fixed paraffin-embedded (FFPE) for targeted sequencing and validation studies, is desirable. Extraction of DNA/RNA from FFPE bone samples can be challenging but has been shown to be feasible as shown in our chondrosarcoma and chordoma genomic landscape manuscripts, although more samples could be made available if samples processing were optimised across the different centres^{18,19}.

2.2.2. MT1-MMP

MT1-MMP is under investigation as a potential biomarker and therapeutic target for OS. Preliminary analyses demonstrated that overexpression of MT1-MMP is present in OS preclinical models and patient tissues; that knockdown of MT1-MMP reduces invasive capability; and that immunohistochemical staining of MT1-MMP on OS biopsy specimens discriminates high grade from low grade OS (Mr Kenneth

Rankin). His group in Newcastle are now currently investigating whether MT1-MMP can be used to detect circulating tumour cells (CTC) through a study involving three centres across the UK. This study has recruited 42 patients to date, and is now starting to optimise retention of potential CTC populations for genomic analysis. The clinical utility of this approach however, requires validation.

A novel molecule targeted to MT1-MMP, BT1718, has been developed by Bicycle Therapeutics in collaboration with Cancer Research UK, and a phase 1 clinical trial commenced in Feb 2018²⁰. All solid cancers with overexpression of MT1-MMP are eligible for study entry, which represents an important opportunity to enrol OS patients into a study that features companion diagnostics as the entry criteria. In addition, preclinical evaluation of an MT1-MMP activated theranostic has commenced in Newcastle University which will include *in vivo* evaluation of MRI enhancement in an orthotopic distal femur mouse bone sarcoma model. If successful this marker could also gain utility to monitor response to treatment. Samples from ICONIC can be used to validate the use of MT1-MMP to identify CTCs, as well as expression in tumour samples to determine suitability for upcoming clinical trials.

2.2.3. Circulating biomarkers

Measurement of circulating tumour cells (CTCs) and circulating tumour or cell free DNA (cfDNA: 'liquid biopsies') have found traction in many cancers: identifying patients with high risk disease, monitoring response to therapy and detecting early relapsed disease. Isolation of CTCs using flow cytometry in OS has been hampered by the lack of a specific antibody²¹. MT1-MMP is a cell surface enzyme overexpressed in the majority of high grade OS and offers one potential method via flow cytometry assessment which is under evaluation at Newcastle. A different approach to isolating CTCs being optimised by Professor Heymann's group in Sheffield (Sarcoma Research Unit) and funded by a BCRT Explorer Grant, uses the Parsortix™ System to enrich for CTCs, which are then isolated and captured using the DEPArray™ System²². The protocols are ready for clinical validation and a pilot study funded by Sarcoma UK to capture CTCs from patients with soft tissue sarcoma has begun recruiting (CIRCUS: A pilot study of CIRCULating tumour cells in patients with soft tissue Sarcoma). Dr Robin Young and Prof. D. Heymann in Sheffield and Saint-Herblain (France) are now currently investigating whether a microfluidic based approach can be used to detect CTCs through a study involving centres across the UK. This study has recruited 25 patients to date. Coordinated collection of blood samples from patients in ICONIC will enable this methodology to be validated efficiently in OS. Copy number variation (CNV) analysis will be used to characterised isolated CTCs.

Professor Flanagan, in collaboration with the Wellcome Sanger Institute has been investigating the utility of cfDNA as a measure of burden of disease. The challenge to this methodology is that patient specific rearrangements identified in the primary tumour are required to quantify cfDNA in the blood. Results from this preliminary analysis are awaited. Another opportunity lies with the potential to use DNA methylation profiles in blood as biomarkers. The potential to collect serial blood samples for multiple analyses within the same cohort of patients provides the opportunity to compare different biomarker assays to better determine their clinical utility.

Patient serum biomarkers such as CRP, albumin and ALP, Hb and ANC:lymphocyte ratio, which are routinely collected, have been investigated as prognostic biomarkers. A novel composite score for sarcoma incorporating five biomarkers (Aarhus composite biomarker score: ACBS) has been shown to be associated with disease specific and overall survival in a retrospective analysis of patients with localised disease²³. This is worthy of prospective validation in a wider cohort of patients.

2.2.4. Immune Oncology

There is a compelling clinical need for improved systemic treatments for patients with OS. Given the efficacy of immune checkpoint inhibitors (CPIs) across more than a dozen different cancer types, immunotherapy has emerged as a promising systemic treatment with potential for durable long-term response. Studies of CPIs in sarcoma have been conducted (e.g., SARCO28), with mixed results showing little to no benefit for patients with OS, and with soft tissue sarcoma responses limited to certain subgroups (e.g. undifferentiated pleomorphic sarcoma). In addition to CPI trials, there are ongoing adaptive T cell therapy trials in a number of sarcoma types, primarily targeting tumour-associated antigens (TAA) including MAGE-A4 and NY-ESO-1. Similarly, the most advanced of these studies is restricted to soft tissue malignancies such as synovial sarcoma, and immunotherapeutic trial activity in OS remains extremely low, with limited prospect of new approved treatments. Despite the promising potential that novel immunotherapies could benefit OS patients, lack of progress in OS can be attributed to multiple factors: (i) the rare nature of the disease and subsequent lack of commercial prioritisation, (ii) small patient/sample sizes available for trials, (iii) lack of relevant OS IO experimental models, and (iv) fundamental gaps in our basic understanding of OS immunobiology. In ICONIC work we aim to address these issues with a step change in the level of collaborative activity, by building on current collaborations and including a novel approach of cross sector public/private working to accelerate the development of novel immunotherapy targets in OS patient samples, as well as a strong focus on closing the gaps in OS immunological understanding.

2.3. Improving systemic therapy and introduction of new treatments

Improvements in systemic therapy for OS are needed urgently. The unmet needs which ICONIC addresses include: addressing variation in delivery of, and response to, standard therapies including chemotherapy and mifamurtide through evaluation of concordance with a national algorithm; identifying factors influencing decision making in specific subgroups such as those aged over 40 or with unresectable primary tumours; stimulating the development of new pilot therapeutic interventions in areas identified by analysis of clinical data on chemotherapy delivery and toxicity, especially for less common subgroups; promotion of access for individual patients to current trials through network impact; provision of the infrastructure shown to be necessary to provide a platform for new agent studies. Examples include: in another rare bone cancer, Ewing sarcoma, the novel multi-arm, multistage (MAMS) study: rEECur (ISRCTN36453794), supported by EU funding to rapidly evaluate treatments at recurrence and compare new agents to standard of care; in lung cancer, driven by the findings from TRACERx, DARWIN 1: Deciphering Afatinib Response and Resistance with INtratour heterogeneity study (NCT02183883).

2.4. Patient Reported Outcomes

The introduction of patient reported outcomes (PROs) into clinical practice is known to improve patient-clinician communication and thus may impact on patient experiences and outcomes²⁴. A sarcoma specific PRO, the Sarcoma Assessment Measure (SAM) has been developed for use in ICONIC with BCRT support. It covers the core domains of physical, emotional, social and financial wellbeing and sexuality so it is a comprehensive reflection of quality of life. The expected utility of SAM includes use as an endpoint in clinical trials as well as for comparing groups of patients over time. Validating the SAM as a longitudinal measure to detect changes in self-reported outcome over time linked to health status may extend its

utility in informing clinicians of changes in health and as a secondary endpoint in future research evaluations.

2.5. Osteosarcoma and routes to diagnosis

Patients with OS often describe prolonged and complex pathways to diagnosis. How this impacts on outcomes and survival is not fully understood, however times to diagnosis have been shown to affect patient experience adversely in more common cancers²⁵. The diagnostic experience of sarcoma patients has been shown to be inferior to those with other more common cancers: sarcoma patients are more likely to report multiple General Practitioner (GP) visits and be dissatisfied with the time taken to see a hospital doctor²⁶⁻²⁸. This requires further investigation and, in particular, examination of the role of the GP and secondary care professionals in the diagnosis of OS. Questionnaire data from patients and GPs together with clinical data will be triangulated to provide a comprehensive description of routes to diagnosis and key time intervals for patients with OS.

2.6. International perspective

EURAMOS-1 was a successful international randomised trial for OS run by four study groups which sought to demonstrate survival benefits from interventions to change the standard of care. Other national study groups joined the EURAMOS core to develop successor trials but no testable question was identified and a recommendation was therefore made for national or cooperative study groups to work internally on hypothesis generating studies, some of which would then be suitable for future wider global evaluation. Although other European groups, notably in France and Italy, are establishing national translational networks for OS, in the UK, to date, no collaborative studies have been developed for OS which have a consistent approach to collection of samples with annotated clinical data. Networking and knowledge sharing between national groups is in operation.

The findings from this trial can contribute to the development of new therapies in collaboration with the newly established FOSTER consortium.

2.7. Benefit to Osteosarcoma Patients

This study will allow us to learn how to be more precise in selecting patients for different treatments, in order to reduce treatment morbidity and improve outcomes. ICONIC investigators will strive to provide an opportunity to all newly diagnosed patients with OS who enter this study to: donate tissue at diagnosis and beyond; contribute clinical information on treatment and outcomes, and participate in the longitudinal validation of a patient reported outcome measure. This will allow us to better understand OS biology to develop biomarkers and understand patient experience, which can be integrated with future treatment strategies.

We expect patients diagnosed with OS in the future to have the opportunity to take part in hypothesis driven clinical trials driven by stratification created from the clinic-biological correlates exposed by the ICONIC investigations. Patient representatives from the NCRI Clinical Studies Group have contributed to the ideas underpinning ICONIC. We will extend patient involvement to encourage a new cadre of patient co-researchers to assist with priority setting and acceptability assessment.

3. STUDY DESIGN

This is a prospective observational cohort study in patients with osteosarcoma (OS). All newly diagnosed OS patients are eligible to take part. Consenting patients are recruited and followed up for at least two years.

The main aim of the study is to establish a platform which supports the recruitment of a consecutive cohort of newly diagnosed OS patients, whose clinical outcomes will be fully annotated and linked to their molecularly characterised biospecimens. The platform will support continuous collection, analysis and interrogation of data to generate hypothesis driven questions quickly for further evaluation. This project extends the Bone Cancer Research Trust tumour banking initiative with funding allocated to provide the infrastructure to optimise collection and storage of tissue samples in the five diagnostic and surgical bone tumour centres in England. It includes the development and validation of clinically relevant biomarkers to identify patients at high risk of relapse and patients suitable for therapeutic studies.

The study will be conducted in two stages. In **Stage 1**, we established the feasibility of patient recruitment and biospecimen collection. In **Stage 2**, correlation with response to chemotherapy and outcome will help develop prognostic biomarkers and provide an opportunity to identify patients for specific therapies. The study will continue these objectives and maintain a platform for development of additional studies and development of therapeutic studies based on the findings from our initial work.

3.1. Study Objectives

3.1.1. Stage 1: Completed

Determined the feasibility of recruiting patients to a national study with high quality clinical data and biospecimen collection to address clinical and biological questions in OS. Its specific research objectives and associated analyses are detailed in [Appendix 4](#) below.

3.1.2. Stage 2

To enrol >350 patients (including those enrolled in Stage 1) and to use the data and samples collected to begin to address the questions listed below.

Following completion of Stage 1 the research objectives of Stage 2 have been amended and adjusted to take into account previous findings:

- Describe variations in management of patients with OS across treatment sites in the UK, including how decisions are reached for tumours being deemed inoperable and how patients contribute to the final decision on surgical operability in terms of what surgery is acceptable.
- Describe current indications for radiotherapy (RT) as part of local therapy
- Describe the impact of variation in management of patients with OS across treatment sites in the UK on patient outcome
- Assess whether tumour margin and response to chemotherapy predict local recurrence in OS
- Use FFPE samples be used to validate WGS findings, identify patients for specific therapies and aid development of novel therapeutic trials
- Use FFPE samples to perform transcriptional and proteomic spatial profiling of OS
- Develop a legacy resource of OS IO data and patient models to accelerate future collaborative research efforts
- Establish novel *in vitro* experimental models of OS (cell lines, organoids, explants) to be used to address biological questions regarding the mechanism of response to chemotherapy and immune-directed therapies
- Create an OS IO data atlas portal
- Assess whether circulating biomarkers can be used as prognostic markers and to predict burden of disease, response to therapy and outcome
- Collect PBMCs from 20 OS patients at UCLH
- Set up an OS imaging repository with NCITA
- Use two methodologies of circulating tumour cell (CTC) detection to isolate and quantify CTCs in a subset of 50 patients undergoing neoadjuvant chemotherapy
- Describe associations between tumour heterogeneity and clonal evolution on chemotherapy response and patient outcome and determine if this can be used to stratify patients for therapy
- Determine if Germline DNA alterations can be confirmed in OS
- Collect WGS samples for the OS population
- Determine if there can be an accelerated target development through partnership with the immune oncology alliance and establishment of OS immune patient models
- Explore whether new candidate immunotherapy drug targets for OS can be discovered based on cellular and genomic analysis
- Assess the value of SAM for longitudinal assessment of patient quality of life and identify predictors of variance in patient reported outcomes (PROs)
- Describe the routes and times to diagnosis for patients with OS, and assess whether this has any impact on patient outcome

3.2. Study Activation

UCL CTC will ensure that all study documentation has been reviewed and approved by all relevant bodies and that the following have been obtained prior to activating the study:

- Health Research Authority (HRA) approval, including Research Ethics Committee approval
- Adoption into NIHR portfolio
- Adequate funding for central coordination
- Confirmation of sponsorship
- Adequate insurance provision

4. SELECTION OF SITES/SITE INVESTIGATORS

4.1. Site Selection

In this protocol study 'site' refers to the hospital where study related activities are conducted.

Sites must be able to comply with:

- Clinical care, follow up schedules and all requirements of the study protocol
- Data collection requirements, including adherence to electronic data capture timelines as per section 11.4
- Biological sample collection, processing and storage requirements
- Monitoring requirements, as outlined in protocol section 14 (Study Monitoring and Oversight)

4.1.1. Selection of Principal Investigator and other investigators at sites

Sites must appoint an appropriate Principal Investigator (PI), i.e. a health care professional authorised by the site to lead and coordinate the work of the study on behalf of the site. Coinvestigators must be trained and approved by the PI. All investigators must be medical doctors and have experience of treating osteosarcoma. The PI is responsible for the conduct of the study at their site and for ensuring that any amendments are implemented in a timely fashion. If a PI plans to take a leave of absence UCL CTC must be informed promptly. For absences greater than three months, or where the PI is no longer able to perform their duties at the site, a new suitable replacement PI must be identified by the site and UCL CTC notified.

UCL CTC may terminate recruitment at a site where a suitable replacement PI has not been identified within three months.

4.1.2. Training requirements for site staff

All site staff must be appropriately qualified by education, training and experience to perform the study related duties allocated to them, which must be recorded on the site delegation log.

CVs for all staff must be kept up to date, signed and dated and copies held in the Investigator Site File (ISF). A current, signed copy of the CV for the PI must be forwarded to UCL CTC upon request.

4.2. Site initiation and Activation

4.2.1. Site initiation

Before a site is activated, the UCL CTC trial team will arrange a site initiation with the site which the PI, and site research team must attend. The site will be trained in the day to day management of the study and essential documentation required for the study will be checked.

Site initiation will be performed for each site by telephone/video conference. Re-initiating sites may be required where there has been a significant delay between initiation and enrolling the first patient.

4.2.2. Required documentation

The following documentation must be submitted by the site to UCL CTC prior to a site being activated by the UCL CTC trial team:

- Site Registration Form (identifying relevant local staff)
- Relevant institutional approvals
- A completed site delegation log that is initialled and dated by the PI (with **all** tasks and responsibilities delegated appropriately)
- A signed and dated copy of the PI's current CV (with documented up to date GCP training, or copy of GCP training certificate)

In addition, the following agreement must be in place:

- A signed site agreement between the Sponsor and the relevant institution (usually an NHS Trust or Health Board)

4.2.3. Site activation

Once the UCL CTC trial team has received all required documentation and the site has been initiated, notification of site activation will be issued to the PI, at which point the site may start to approach patients.

Following site activation, the PI is responsible for ensuring:

- adherence to the most recent version of the protocol and Laboratory Manual which provides detailed instructions on biological sample collection, processing, storage and shipping
- all relevant site staff are trained in the protocol requirements
- appropriate recruitment and medical care of patients in the study
- timely completion of eCRFs (including assessment of all adverse reactions related to study procedures)
- prompt notification and assessment of all serious adverse reactions related to study procedures

5. INFORMED CONSENT

Sites are responsible for assessing a patient's capacity to give informed consent.

Sites must ensure that all patients have been given the appropriate current approved version of the patient information sheet(s), are fully informed about the study and have confirmed their willingness to take part in the study by signing the appropriate current approved consent form.

Sites must assess a patient's ability to understand verbal and written information in English and whether or not an independent interpreter or NHS approved translator would be required to ensure fully informed consent. If a patient requires an interpreter and none are available prior to consent, and for the duration of the potential participant's time on the study, the patient should not be considered for the study.

The PI, or, where delegated by the PI, other appropriately trained site staff, are required to provide a full explanation of the study to each patient prior to study entry. During these discussions, the current approved patient information sheet for the study should be discussed with the patient.

Patients should be given adequate time to consider and discuss participation in the study. However, patients may consent on the same day they are given the information sheet if this is more convenient for them. In such cases, a member of the research team at the hospital must phone the patient in the following days to confirm that they are still willing to participate in the study.

Written informed consent on the current approved versions of the consent forms for the study must be obtained before any study specific procedures are conducted. The discussion and consent process must be documented in the patient medical notes.

Site staff are responsible for:

- checking that the current approved versions of the patient information sheets and consent forms are used
- giving the patient a copy of the patient information sheet
- checking that information on the consent form is complete and legible
- checking that the patient has initialled **all** relevant sections of the consent form and signed and dated the form
- checking that an appropriate member of staff has countersigned and dated the consent form to confirm that they provided information to the patient giving the patient a copy of their signed consent form
- checking that an appropriate member of staff has made dated entries in the patient's medical notes relating to the informed consent process (i.e. information given, consent signed etc.)
- following registration, adding the patient's study number to all copies of the consent form, which should be filed in the patient's medical notes and investigator site file

The right of the patient to refuse to participate in the study without giving reasons must be respected. All patients are free to withdraw at any time. Also refer to section 15 (withdrawal of patients).

5.1. Additional information for patients under 16

The person with parental responsibility or legal guardianship of the child must be informed of all aspects of the study. The child must also be informed about the study to the extent compatible with their understanding. The same information must be provided to children of all ages, however the level of detail is age appropriate.

Information sheets are available for the following age groups:

- 06 – 09 years old
- 10 – 12 years old
- 13 – 15 years old

There is also an information sheet available for parents/legal guardians.

The patient and parent/legal guardian should be given adequate time to consider and discuss participation in the study. However, the patient and parent/legal guardian may consent on the same day they are given the information sheet if this is more convenient for them. In such cases, a member of the research team at the hospital must phone the patient and parent/legal guardian in the following days to confirm that they are still willing to participate in the study.

Written informed consent on the current approved version of the parent/legal guardian consent form for the study must be obtained before any study specific procedures are conducted. If capable the child may give assent by signing and personally dating the informed consent form, in addition to the parent or legal guardian. The discussion and consent process must be documented in the patient medical notes.

A child's refusal to participate in the study must be respected.

5.2. Additional information for adult patients lacking capacity to consent

- **Section 5.2 is NOT to be applied at any Scottish or Northern Irish ICONIC sites.**

It is recognised that there are several genetic syndromes which cause both osteosarcoma and learning disabilities which may mean a patient does not have the capacity to consent. Where possible, these patients should be given the opportunity to participate in the study.

The person with duty of care for a patient lacking capacity must be informed of all aspects of the study. Where appropriate, the patient should also be informed about the study to the extent compatible with their understanding (for example one of the child patient information sheets could be used that is compatible with the patient's understanding).

The person with duty of care should be given the consultee information sheet. The person with duty of care, and the patient (where appropriate) should be given adequate time to consider and discuss participation in the study. However, confirmation of opinion that the patient would have no objection to taking part may be given on the same day as the information sheet if this is more convenient. In such cases, a member of the research team at the hospital must phone the person with duty of care in the following days to confirm that the patient is still willing to participate in the study.

Written confirmation of opinion that the patient would have no objection to taking part in the study will be collected on the current approved version of the consultee declaration form for the study, and must be obtained before any study specific procedures are conducted. If capable the patient may give assent by signing and personally dating the consultee declaration form, in addition to the person with duty of care. The discussion and process of confirming the patient would have no objection to taking part in the study must be documented in the patient notes.

A patient's refusal to participate in the study must be respected.

5.3. Remote consent

[Appendix 1](#) gives guidance outlining the process for sites to follow if it is not possible for consent to be taken in person. All steps performed need to be documented clearly and filed in patient notes accordingly.

6. SELECTION OF PATIENTS

6.1. Screening Log

A screening log must be maintained and appropriately filed at site. Sites should record all patients identified with newly diagnosed osteosarcoma and the reasons why they were not registered in the study if this is the case. The log must be sent to UCL CTC when requested.

6.2. Patient Eligibility

Queries in relation to the eligibility criteria should be addressed prior to registration. Patients are eligible for the study if all the inclusion criteria are met and none of the exclusion criteria apply.

Patient eligibility must be confirmed by an investigator who is suitably qualified and who has been allocated this duty, as documented on the site staff delegation log, prior to registering the patient. Confirmation of eligibility must be documented in the patient medical notes and on the registration eCRF.

If a potentially eligible patient has a second primary OS, UCL CTC must be contacted before the patient is approached, so that eligibility for the study can be confirmed.

Patients must give written informed consent before any study specific investigations may be carried out. Refer to section 9.1 (Preregistration Assessments) for the list of assessments and procedures required to evaluate the suitability of patients prior to entry.

Patients who do not wish to complete the PROs but are willing to donate blood and/or tissue samples are eligible for the Study.

6.2.1. Inclusion criteria

- New histological diagnosis of osteosarcoma or in the absence of osteoid seen on biopsy, pathology and imaging supportive of a diagnosis of osteosarcoma.

(It is well recognised that some patients may present with features suggestive of osteosarcoma (under 40 years, radiological abnormality compatible) but in whom no osteoid is detected in needle biopsy. Although categorised as spindle cell tumour of bone, such patients are usually treated in an identical approach to osteosarcoma. A definite diagnosis of osteosarcoma is then often possible after surgery when the entire resection specimen is available.)

- Written informed consent of patient and/or parent/legal guardian or written confirmation of opinion of person with duty of care where applicable.

6.2.2. Exclusion criteria

- Diagnosis more than four months prior to registration

7. REGISTRATION PROCEDURES

Patient registration will be performed via a remote electronic data capture system (MACRO) hosted by UCL CTC. Please refer to the registration instructions provided in the ICONIC Database User Manual for sites. Patients must be confirmed to be eligible and have given consent prior to registration.

Site staff responsible for patient registration must request access to the eCRF database by completing the delegation log and their contact details on the site database user access form. Access to the database and instructions are provided by UCL CTC.

Note that patient initials, age at study entry, details of the Sarcoma MDT site and date of diagnosis are required to register a patient.

Upon registration confirmation from UCL CTC a study number will be assigned to the patient. This will be in the registration confirmation email which UCL CTC will send to the person registering the patient.

The study number must be recorded in the patient notes.

CONTACT DETAILS	
ICONIC Trial Coordinator:	020 7679 9878
ICONIC email:	ctc.iconic@ucl.ac.uk
ICONIC Remote Db Access:	https://rde.ctc.ucl.ac.uk/
UCL CTC office hours: 09:00 to 17:00 Monday to Friday excluding Bank Holidays (UK time)	

Once a patient has been registered onto the study they must be provided with the following:

- A copy of their signed consent form and patient information sheet(s).

After registration into the trial, the patient's general practitioner (GP) should be informed of the patient's involvement in the trial by the site completing and sending the completed GP letter, GP early diagnosis form and information sheet.

8. STUDY TREATMENT

No treatments are specified by this protocol. Patients should be treated according to national guidelines including *UK guidelines for the management of bone sarcomas*²⁹ and applicable current and future NICE guidance.

9. STUDY ACTIVITIES

Please also see Schedule of Events tables in [Appendix 2](#)

9.1. Preregistration

The following is required to evaluate the suitability of patients for the study:

- Histological confirmation of osteosarcoma, including date

The following patient information is also required in order to register the patient:

- Age at time of registration
- Sex

9.2. Baseline

9.2.1. Baseline: Clinical Data

The following data should be collected from routine assessments carried out within 28 days of registration:

Medical History

- History of previous cancer
- Relevant previous medical conditions, procedures and treatments
- Current medical conditions, signs and symptoms
- Cancer signs and symptoms
- Duration of symptoms
- Causative risk factors (family history, previous radiotherapy, Paget's disease etc.)
- Smoking status

Baseline characteristics

- Physical examination (height and weight)
- Assessment of WHO performance status (patients ≥ 16) or Lansky performance status (see [Appendix 3](#))
- Haematology: haemoglobin, ANC, lymphocytes, platelets
- Biochemistry: CRP, albumin, ALP, LDH, creatinine

Primary Tumour: Diagnostic imaging

- Date and type of scan and hospital where this was performed
- Site, size and location on bone of primary tumour
- Description of findings (intraarticular, pathological fracture, involvement of neurovascular bundle, presence of skip metastases etc.)

Primary tumour: Biopsy

- Date of biopsy and hospital where this was performed
- Tumour grade (assessed using TNM Classification for Osteogenic Sarcoma 15/01/2018) and histological subtype

Primary tumour: Surgery (if already performed)

- Date and type of surgery and hospital where this was performed
- Histological subtype
- Resection margins

Diagnostic staging: Imaging

- Date and type of imaging and hospital where this was performed
- Location, number and size of metastases, if applicable

Treatment plan

- Name and date of MDM where patient discussed
- Type of planned surgical operation or reason if surgery not to be performed (if known)
- Chemotherapy regimen, if planned or reason if no chemotherapy to be given
- Details on plans for radiotherapy or other care
- Adverse reactions: assessed only in relation to study procedures, e.g. venepuncture, biopsy for research when applicable.

9.2.2. Baseline: PROs

Patients aged 13 or over should complete the PROs including Sarcoma Assessment Measure (SAM), EORTC-QLQ-C30, TESS and GRC within 2 weeks after registration and prior to starting treatment (if possible). The SAM booklet (Sarcoma Assessment Measure (SAM), EORTC-QLQ-C30, TESS and GRC) can be completed over the phone as well as by post or in person.

9.2.3. Baseline: Routes to Diagnosis

The patient routes to diagnosis questionnaire should be given to the patient or parent/legal guardian at baseline only. It must be completed in person and not undertaken over the phone or posted. It takes around 20 minutes to complete and should be returned to site for data entry.

- Patients aged 13 or over should complete the Patient routes to diagnosis questionnaire within three months of registration.
- For patients aged less than 13, the parents or legal guardians should complete the Patient routes to diagnosis questionnaire within three months of registration.

The GP routes to diagnosis questionnaire and information sheet should be sent (by email or post) to the patient's GP at baseline and should be returned to site for data entry. If not returned to the site within four weeks the GP should be sent one reminder.

9.2.4. Baseline: Research Samples

(SEE [SECTION 10](#) FOR SUMMARY)

All patients

- Whole blood sample for germline DNA for the ECMC GCLP Facility (preferably collected at study entry, but may be collected at any point during the study)
- Whole blood sample for ctDNA, DNA methylation profiling/future research for the ECMC GCLP Facility (**optional**)
 - NB if patient has started chemotherapy prior to study entry this sample may still be collected
 - If the baseline ctDNA sample is missed for any reason, please continue with the on treatment, end of treatment and follow up samples.
- Surgery site to ship diagnostic FFPE tissue to RNOH (coordinated by UCL CTC)
- Collect information about frozen tumour samples (where sample stored, reason sample not taken if applicable)

Patients receiving neoadjuvant chemotherapy

- Whole blood samples for CTCs, collected prior to the start of chemotherapy

Patients on PBMC substudy (UCLH site only)

- Whole blood samples for PBMCs, collected prior to the start of neoadjuvant chemotherapy.

9.3. During Treatment

If patient does not receive treatment for any reason, skip to section 9.5 ([Follow up](#)).

The reason why no treatment was given should be documented.

9.3.1. During Treatment: Clinical Data

During treatment, data should be collected at the end of neoadjuvant chemotherapy, at surgery, at the end of adjuvant chemotherapy, after radiotherapy and at the end of treatment.

The following details should be collected at the end of neoadjuvant chemotherapy:

- Details of chemotherapy given, including drug doses, dates and reason(s) for reductions, delays or discontinuation
 - If neoadjuvant chemotherapy not given, reason should be documented
- Disease status (if assessment carried out)
- Adverse reactions: assessed only in relation to study procedures, e.g. venepuncture, biopsy for research, when applicable

The following details should be collected prior to surgery:

- WHO performance status (patients ≥ 16) or Lansky performance status

- Blood Biochemistry: CRP, albumin, ALP, LDH, creatinine
- Haematology: Hb, ANC, lymphocytes, platelets
- Preoperative imaging
 - Date of MRI
 - Date of CT, if performed
 - Date and type of other imaging, if applicable
 - Disease status

The following surgical details should be collected following surgery for ALL patients on the Surgery eCRF:

- Date and time of surgery
- Hospital where surgery performed
- Type of surgery (amputation, limb sparing etc.)
- Details of surgical procedure (which anatomical structures removed, use of antibiotics, details on blood loss, whether central line in situ, type of reconstruction, surgical margin assessment etc.)
- ASA classification
- Postoperative complications (wound complications, dehiscence, infection or haematoma requiring return to theatre, thromboembolism, deep implant infection etc.)
- **If surgery was planned but not done the reason should be documented.**

Pathology

- Maximum dimension on pathology report
- Margins: bone margin, narrowest margin in mm
- Lymphovascular invasion
- Response to chemotherapy, % necrosis
- Was a fresh/frozen specimen stored?
 - If so, where?
 - If not, why not?
- Adverse reactions: assessed only in relation to study procedures, e.g. venepuncture, biopsy for research, when applicable

The following details should be collected at the end of adjuvant chemotherapy:

- Details of chemotherapy given, including drug doses, dates and reason(s) for reductions, delays or discontinuation
- If adjuvant chemotherapy not given, reason
- Was mifamurtide given? If not, reason why
- Disease status (if imaging assessment carried out)
- Adverse reactions: assessed only in relation to study procedures, e.g. venepuncture

The following details should be collected after radiotherapy (if given):

- Details of radiotherapy, including indication (primary, palliative or both), modality, technique, dates, doses and fractionation
- Adverse reactions: assessed only in relation to study procedures

If patients undergo thoracotomy at any time, please provide:

- Date, place of surgery;
- type of resection; side of resection; no of lesions resected;
- was metastatic disease verified histologically?
- no. of verified lesions; resection margins
- Was sample stored for research?
 - if so, where is it stored?

9.3.2. During Treatment: Research Samples

All patients

- Whole blood sample for ctDNA, DNA methylation profiling/future research for the ECMC GCLP Facility (**optional**)
 - For patients receiving MAP chemotherapy collect Pre cycle 2 day 21 methotrexate (ideally up to 3 to 7 days prior/14 days post cycle 2) – or pre surgery if this is not possible
 - For patients receiving AP or other chemotherapy, collect pre cycle 3 (ideally up to 3 to 7 days prior and 14 days post cycle 2) – **or pre surgery** if this is not possible
 - If the baseline ctDNA sample was missed for any reason, please continue with the On treatment, End of treatment and Follow up samples.

Patients receiving neoadjuvant chemotherapy

- Whole blood sample for CTCs
 - Only to be taken if the baseline CTC sample was collected.
 - For patients receiving MAP chemotherapy collect Pre cycle 2 day 21 methotrexate (ideally up to 3 to 7 days prior/14 days post cycle 2) – or pre surgery if this is not possible
 - For patients receiving AP or other chemotherapy, collect pre cycle 3 (ideally up to 3 to 7 days prior and 14 days post cycle 2) – or pre surgery if this is not possible

Patients on PBMC substudy (UCLH site only)

- Whole blood samples for PBMCs, collected prior to surgery

9.4. End of Treatment

- **For the purpose of clinical data, sample and PROs collection End of Treatment is defined as end of chemotherapy, and not end of treatment with Mifamurtide**

9.4.1. End of Treatment: Clinical data

The following details should be collected at the end of treatment:

- Total number of cycles of chemotherapy and doses
- Total number of mifamurtide cycles
- If terminated early, provide reason
- Imaging performed, date, modality, result
- Blood Biochemistry: CRP, albumin, ALP, LDH, creatinine
- Haematology: Hb, ANC, lymphocytes, platelets
- Disease status
- Adverse reactions: assessed only in relation to study procedures, e.g. venepuncture, biopsy for research.

9.4.2. End of Treatment: PROs

- Patients aged 13 or over should complete the PROs (SAM, EORTC QLQ-C30, TESS, & GRC) at the end of treatment.

9.4.3. End of Treatment: Research Samples

All patients

- Whole blood sample for ctDNA, DNA methylation profiling/future research for the ECMC GCLP Facility (**optional**)
- Surgery site to ship FFPE tissue from the surgical resection to RNOH (coordinated by UCL CTC)

Patients on PBMC substudy (UCLH site only)

- Whole blood samples for PBMCs

9.5. Follow up

9.5.1. Follow up: Clinical Data

Follow up visits should take place, at least annually, according to routine practice, on all patients including those who did not receive treatment. The following details should be collected:

- Disease status
- Survival status
- Details on any post-operative complications (e.g. deep implant infection)
- Adverse reactions: assessed only in relation to study procedures, e.g. venepuncture, biopsy for research, when applicable

Please continue completing all follow up forms after patient has relapse/metastasis.

9.5.2. Follow up: PROs

- Patients aged 13 or over should complete the PROs including SAM, EORTC-QLQ-C30, TESS, GRC annually from registration until relapse.

9.5.3. Follow up: Research Samples

All patients

- Whole blood sample for ctDNA, DNA methylation profiling/future research for the ECMC GCLP Facility collected at clinic visit during follow up no more than 6 monthly (**optional**).

9.6. Relapse (local recurrence or metastases)

9.6.1. Relapse: Clinical Data

The following data should be collected on confirmation of relapse (local recurrence or metastases) and at every subsequent relapse:

- Site(s) of relapse (local, distant etc.)
- Treatment received (surgery for local recurrence or metastases, systemic therapy, radiotherapy etc.)
- Adverse reactions: assessed only in relation to study procedures, e.g. venepuncture, biopsy for research.

Please continue completing all follow up forms after patient has relapse/metastasis.

9.6.2. Relapse: Research Samples

All patients

- Whole blood sample for ctDNA, DNA methylation profiling/future research for the ECMC GCLP Facility (**optional**)
- If the baseline optional ctDNA sample was missed for any reason, please continue with the on treatment, end of treatment and follow up samples
- Tumour tissue collected as part of standard of care, or for research (**optional**) (FFPE and fresh/frozen according to local policy, if possible, and stored locally at surgical site: collection coordinated by UCL CTC)

Patients receiving neoadjuvant chemotherapy

- Blood samples for CTCs (**only** if the baseline and pre surgery samples have been collected).

Patients on PBMC substudy (UCLH site only)

- Whole blood samples for PBMCs

10. EXPLORATORY BIOLOGICAL STUDIES

The following sections provide an overview of sample collection. For details of sample collection, processing and shipping, refer to the **ICONIC Laboratory Manual**.

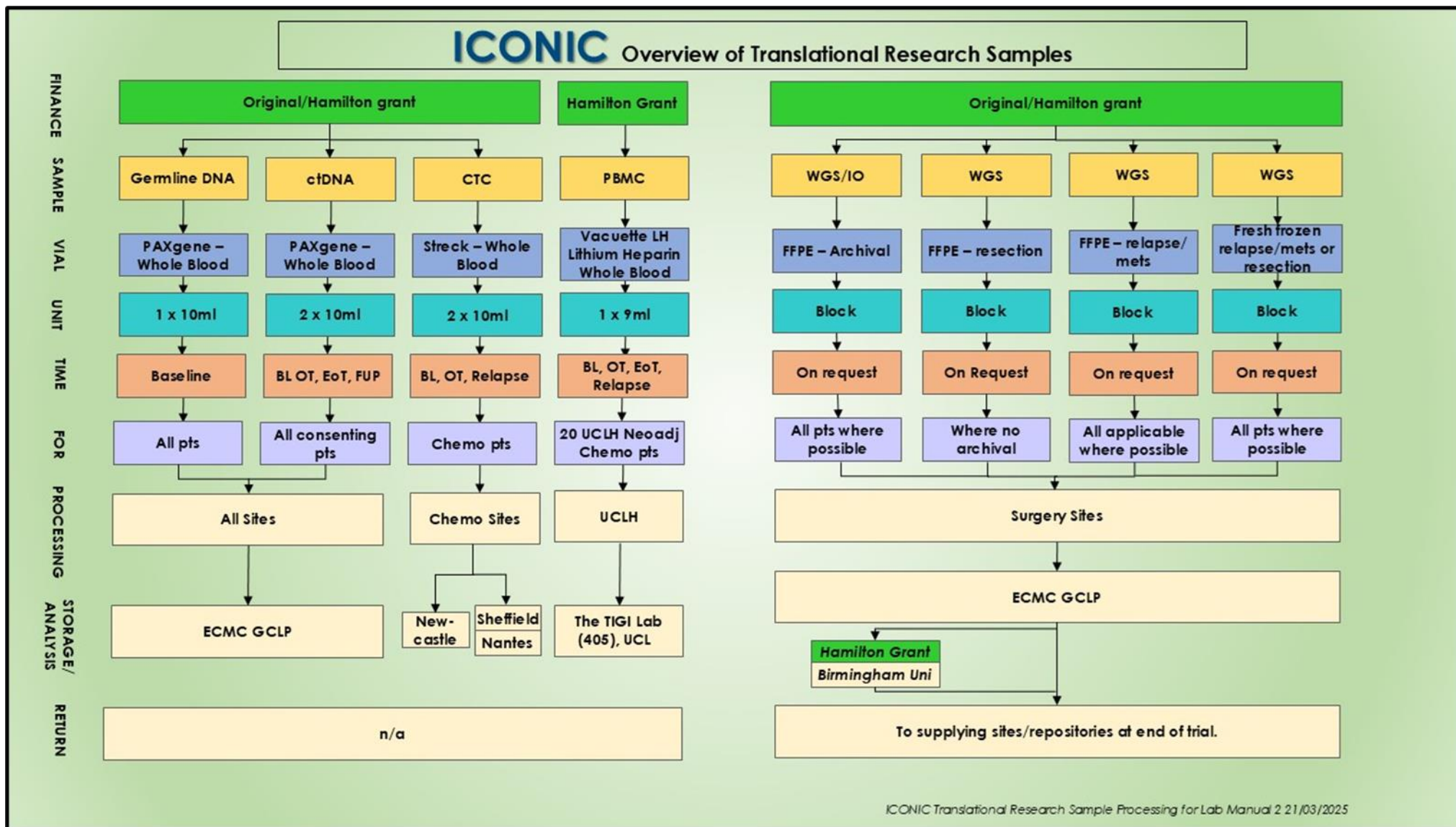
Below are two tables outlining biological sample collection for patients receiving either neoadjuvant chemotherapy, or surgery with or without adjuvant chemotherapy. The following sections give further details on processing, collection and storage.

SCHEDULE: PATIENTS RECEIVING SURGERY ALONE +/- ADJUVANT CHEMOTHERAPY	Within 28 days after registration	Surgery	End of Treatment	Follow up	Relapse (local recurrence or metastases)
Whole blood sample for ctDNA (optional for all patients) for the ECMC GCLP Facility	X		X	X ¹	X
FFPE tumour tissue for RNOH	X	X			X ²
Provide information on frozen tumour tissue ³	X	X	X		X ²
Frozen tissue samples sent on request	X	X			X

1. To be collected at clinic visit during follow up no more than 6 monthly.
2. Collect if patient consents to this optional biopsy (if not already collected as part of SOC)
3. Where sample stored, reason sample not taken if applicable

SCHEDULE: PATIENTS RECEIVING NEOADJUVANT CHEMOTHERAPY	Baseline: within 28 days after registration	During Treatment		End of Treatment	Follow up	Relapse (local recurrence or metastases)
		End of Neoadjuvant chemotherapy	Surgery			
Whole blood sample for germline DNA for the ECMC GCLP Facility	X					
Whole blood sample for CTCs	X	X	(X) ¹			X
Whole blood sample for ctDNA for the ECMC GCLP Facility (optional for all patients)	X	X	(X) ¹	X	X ²	X
PBMCs (UCLH only)	X	X	(X) ¹	X		X
FFPE tumour tissue for RNOH	X		X			X ³
Provide information on frozen tumour tissue ⁴	X		X	X		X ³
Frozen tissue samples sent on request	X		X			X

1. Can collect before primary resection surgery if not taken at end of neoadjuvant treatment
2. To be collected at clinic visit during follow up no more than 6 monthly.
3. Collect if patient consents to this optional biopsy (if not already collected as part of SOC).
4. Where sample stored, reason sample not taken if applicable



10.1. FFPE tumour tissue samples

FFPE samples should be obtained for all patients and shipped to the Royal National Orthopaedic Hospital (RNOH). These will be used to optimise processes to enable DNA/RNA extraction from samples to perform molecular targeted sequencing and copy number analysis (CNA), and to validate findings from reported and ongoing WGS studies.

The collection of samples will be coordinated centrally by UCL CTC.

10.1.1. FFPE Collection time points

The following FFPE samples will be collected:

- Archival FFPE tissue blocks from previous biopsy or surgery
- FFPE tissue blocks from routine surgery taking place after registration
- FFPE collected at relapse/mets as part of standard of care, or for research (**optional**) (see section 10.3 below).

10.1.2. FFPE Processing

All samples should be fixed and embedded according to local practice.

For routine samples, sites should obtain at least one and ideally two FFPE tumour tissue blocks from a previous biopsy or surgery.

Review the pathology report and select blocks as follows:

- Biopsy cores: Choose cores that have the highest tumour tissue content.
- Surgical resection tissue: Choose blocks containing the most tumour tissue.

10.1.3. FFPE Shipping

Archival samples should be shipped on request from UCL CTC.

10.1.4. FFPE Return

All FFPE samples will be stored at RNOH. During the Study sites may request return of routine samples if required for clinical purposes but would be requested to return the samples to RNOH after this.

At the end of the Study all routine FFPE samples will be returned to site.

Samples taken for research (i.e. those not clinically indicated) will not be returned to sites.

10.2. Fresh/Frozen tumour specimens

Information will be captured about routine collection of frozen tumour specimens, at baseline, surgical resection of primary tumour or metastases and on relapse/local recurrence.

- Fresh/frozen tumour specimens should be shipped on request from UCL CTC.
- At the end of the Study all routine fresh/frozen tumour specimens will be returned to site.

10.3. New biopsy samples (Fresh/Frozen and FFPE) at relapse or metastases

A new biopsy, FFPE and fresh/frozen according to local policy if possible, should be obtained at relapse/resection of local recurrence or metastases for the study if not done as part of standard of care. This is optional for patients.

The collection of samples will be coordinated centrally by UCL CTC.

For new biopsies for research, two cores should be obtained. One core should be fresh frozen in liquid nitrogen according to local policy and one core should be formalin fixed paraffin embedded.

If it is not possible to obtain two cores, then one is acceptable and should be fresh/frozen according to local policy.

- The fresh/frozen sample should be stored at site at -80°C until requested for analysis.

10.4. Whole Blood samples for Circulating Tumour Cells

Blood samples will be collected for analysis of circulating tumour cells (CTCs) in patients receiving neoadjuvant chemotherapy.

Samples sent to Newcastle University will be used for ongoing validation of the MT1-MMP method of CTC isolation through the use of flow cytometry and subsequent genetic characterisation.

Samples sent to the University of Sheffield will be used to optimise a new method for isolating CTCs. The procedure uses the Parsortix™ System to enrich for CTCs, which are then isolated and captured using the DEPArray™ System²².

The protocols are ready for clinical validation and a pilot study funded by Sarcoma UK to capture CTCs from patients with soft tissue sarcoma has begun recruiting. Blood samples from patients in ICONIC will enable this methodology to be validated in OS.

10.4.1. CTC Collection time points

CTC samples will be collected for at least 50 neoadjuvant chemotherapy patients .

Do not collect these samples if there is no plan for the patient to have neoadjuvant chemotherapy.

Samples should be collected at the following timepoints (at the same time as routine bloods are taken):

- Baseline (prior to start of neoadjuvant chemotherapy)
- During treatment (**only if the baseline sample has been collected**):
 - For patients receiving **MAP** chemotherapy collect Pre cycle 2 day 21 methotrexate (ideally up to 3 to 7 days prior/14 days post cycle 2) – or pre surgery if this is not possible
 - For patients receiving **AP or other** chemotherapy, collect pre cycle 3 (ideally up to 3 to 7 days prior and 14 days post cycle 2) – or pre surgery if this is not possible
- At relapse (local recurrence or metastases) and **only if the first two samples have been collected.**

10.4.2. Processing and shipping

- Whole blood should be collected into 2 x 10 mL Streck tubes and inverted 10 times to mix.
- Samples must be shipped on the day they are collected, using the shipping materials provided.
- One sample should be shipped to Newcastle University and the other to the University of Sheffield.

10.5. Whole blood samples for ctDNA, methylation profiles and future research

Whole blood samples for future research including measurement of cell free DNA and analysis of methylation profiles may be collected from all patients who have consented to this. **These samples are optional.**

10.5.1. ctDNA Collection time points

Samples from all patients consenting to this should be collected at the following timepoints (at the same time that routine bloods are taken):

- Baseline (prior to start of neoadjuvant chemotherapy if applicable)
- During treatment:
 - For patients receiving **MAP** chemotherapy collect Pre cycle 2 day 21 methotrexate (ideally up to 3 to 7 days prior and 14 days post cycle 2) – or pre surgery if this is not possible
 - For patients receiving **AP or other** chemotherapy, collect pre cycle 3 (ideally up to 3 to 7 days prior and 14 days post cycle 2) – or pre surgery if this is not possible
- End of chemotherapy treatment (**NOT** end of mifamurtide treatment if applicable)
- At clinic visit during follow up no more than 6 monthly.
- At relapse (local recurrence or metastases)

10.5.2. ctDNA Processing and shipping

Whole blood should be collected into 2 x 10 mL BD Vacutainer® PAXgene ccfDNA blood collection tubes and inverted 10 times to mix. Samples must be shipped to the ECMC GCLP Facility on the day they are collected, using the shipping materials provided. For details please refer to the ICONIC Laboratory Manual.

10.6. Whole blood samples for germline DNA

Blood samples for germline DNA will be collected from all patients and used for WGS and targeted sequencing.

10.6.1. Germline DNA Collection time points

One sample should preferably be collected at baseline (study entry) and can be collected at the same time as routine bloods are taken, if not at baseline, at any point after consent until the end of the trial.

10.6.2. Germline DNA Processing and shipping

Whole blood should be collected into a 10 mL BD Vacutainer® PAXgene ccfDNA blood collection tube and inverted 10 times to mix.

Samples must be shipped to the ECMC GCLP Facility on the day they are collected, using the shipping materials provided.

10.7. Whole blood samples for PBMCs (UCLH site only)

Blood samples for PBMCs will be collected from 20 UCLH patients.

10.7.1. PBMC Collection time points

- Baseline/pre neoadjuvant chemotherapy
- Presurgery
- End of Treatment (end of adjuvant chemo)
- Relapse (if applicable)

10.7.2. PBMC Processing and shipping

Whole blood should be collected into a Vacuette™ LH Lithium Heparin blood collection tube: preferably 10 mL but 9 mL is acceptable.

Samples must be delivered to the TIGI Lab (405), at the UCL Cancer Institute, Paul O’Gorman Building, 72 Huntley Street, London, WC1E 6DD on the day of collection.

11. DATA MANAGEMENT AND DATA HANDLING GUIDELINES

Data will be collected from sites using an eCRF (electronic case report form) created and maintained by UCL CTC. Data must be accurately entered into the eCRFs and must be verifiable from source data at site. Examples of source documents are hospital records which include patient medical notes, laboratory and other clinical reports, etc.

11.1. Entering data into the eCRF

The eCRF must be completed by site staff who have been appropriately trained, are listed on the site staff delegation log and authorised by the PI to perform this duty. Each authorised staff member will be issued with their own unique login details for the eCRF by UCL CTC, and a list of current users at each site will be maintained by UCL CTC. Site staff must never share their login details with other staff as the eCRF audit trail will record all entries/changes made by each user. The PI is responsible for the accuracy of all data reported in the eCRF.

The use of abbreviations and acronyms should be avoided.

11.2. Corrections to eCRF Forms

Where necessary corrections can be made by site staff to data on the eCRF, as long as the eCRF has not been locked or frozen by UCL CTC. The eCRF audit trail will record the original data, the change made, the user making the change and the date and time. Site staff should contact UCL CTC if changes need to be made to a locked or frozen eCRF.

11.3. Missing Data

To avoid the need for unnecessary data queries, fields should not be left blank on the eCRF. If data is unavailable, please refer to the ICONIC Database Manual for sites for information on how to indicate that data is “Not Done”, “Not Applicable”, “Not Available” or “Not Known” (only use if every effort has been made to obtain the data).

11.4. Timelines for Data Entry

The relevant eCRFs must be completed as soon as possible after a patient’s visit. Registration eCRFs must be completed for a patient to be registered onto the study. All other eCRFs must be completed within 7 days of the patient being seen.

Sites who persistently do not enter data within the required timelines may be suspended from recruiting further patients into the study by UCL CTC and this may trigger a monitoring visit. See section 14.2 (‘Triggered’ Onsite/Remote Monitoring) for details.

11.5. Data Queries

Data entered onto the eCRF will be subject to some basic checks at the time of entry, and any discrepancies will be flagged to the user in the form of a warning. The data can be corrected immediately, or where this is not possible, the warning can be saved and the data amended at a later stage.

Further data review will be carried out at UCL CTC and queries raised where necessary. Further guidance on the process for handling data queries can be found in the ICONIC Database Manual for sites.

There may be times when data queries require a rapid response. UCL CTC will contact sites if this is the case and provide as much notice as possible.

12. SAFETY REPORTING

12.1. Definitions

The following definitions have been adapted from the Medicines for Human Use (Clinical Trials) Regulations 2004 (SI 2004/1031) and subsequent amendments, ICH E2A “Clinical Safety Data Management: Definitions and Standards for Expedited Reporting” and ICH GCP E6.

This is a low risk prospective observational cohort study.

Adverse Reactions (AR)

All untoward and unintended events causally related to a ‘Study Procedure’; where a causal relationship between a ‘Study Procedure’ and an event is at least a reasonable possibility, i.e. the relationship cannot be ruled out.

Serious Adverse Reactions (SAR)

SARs are an adverse reaction that meets any of the following criteria:

- results in death
- is life-threatening (the term “life-threatening” refers to an event in which the patient was at risk of death at the time of the event. It does not refer to an event that hypothetically might have caused death if it were more severe)
- requires inpatient hospitalisation or prolongs existing hospitalisation
- results in persistent or significant disability/incapacity
- is otherwise medically significant (e.g. important medical events that may not be immediately life-threatening or result in death or hospitalisation but may jeopardise the patient or may require intervention to prevent one of the other outcomes listed above)

Related & Unexpected SARs

A serious adverse reaction, the nature or severity of which is **not consistent** with the applicable Study Procedure.

Study Procedure means the blood sampling procedure and/or biopsy procedure for the purposes of obtaining research samples within the study.

12.2. Serious Adverse Reactions (SARs)

SARs are not anticipated in this study however SARs that are attributable to the study procedures will be reported immediately to the UCL CTC and documented on the study database.

12.3.1 Reporting of Serious Adverse Reactions (SARs)

As this is an observational cohort study where patients follow their normal clinical pathways, and the study introduces two procedural interventions (blood sampling and biopsies), the PI, or other delegated site investigator should monitor each participant at each visit and only report to UCL CTC events that are serious and related (i.e. a SAR) to the study procedure.

Causality

The PI, or other delegated site investigator, must perform an evaluation of causality for each SAR. UCL CTC will consider events evaluated as related to be adverse reactions.

- Related (reasonable possibility) to a study procedure
- Not related (no reasonable possibility) to a study procedure

Severity

Severity of each event must be determined by using the Common Terminology Criteria for Adverse Events (CTCAE) v5 as a guideline, wherever possible. The criteria are available online at:

https://ctep.cancer.gov/protocoldevelopment/electronic_applications/docs/CTCAE_v5_Quick_Reference_5x7.pdf

In those cases where the CTCAE criteria do not apply, severity should be coded according to the following criteria:

- 1 = Mild (awareness of sign or symptom, but easily tolerated)
- 2 = Moderate (discomfort enough to cause interference with normal daily activities)
- 3 = Severe (inability to perform normal daily activities)
- 4 = Life threatening (immediate risk of death from the reaction as it occurred)
- 5 = Fatal (the event resulted in death)

For the purposes of the study, SARs of all grades should be reported.

All SARs that occur between the start of the first study procedure and 15 days post the last study procedure must be submitted electronically within **24 hours** of observing or notification/occurrence of the event, using the study specific SAR Report.

All sections on the SAR Report must be completed. If the SAR report **is not sent within 24 hours** to UCL CTC, the circumstances that led to this must be detailed in the SAR Report to avoid unnecessary queries.

Completed SAR Reports must be submitted to UCL CTC within 24 hours of becoming aware of the event

Email: CTC.iconic@ucl.ac.uk

SAR Follow Up Reports

All SARs must be followed up until resolution and until there are no further queries.

Sites must ensure any new and relevant information is provided promptly. If the reaction term changes or a new reaction is added, the causality must be re-assessed by an Investigator.

SAR Processing at UCL CTC

On receipt of the SAR Report, UCL CTC will check for legibility, completeness, accuracy and consistency. There are no expected SARs for the study, therefore all will be considered Related and Unexpected Serious Adverse Reactions.

The CI, or their delegate (e.g. a clinical member of the TMG) will be contacted to review the SAR and to perform an evaluation of causality on behalf of UCL CTC.

12.3. Related and Unexpected Serious Adverse Reaction

If the event is evaluated as a related and unexpected Serious Adverse Reaction, UCL CTC will submit a report to the REC within the required timeline. For this study, all SARs will be considered as unexpected.

Wherever possible, evaluations of causal relationship by both the site and the Sponsor's clinical reviewer will be reported.

13. INCIDENT REPORTING AND SERIOUS BREACHES

13.1. Incident Reporting

Organisations must notify UCL CTC of all deviations from the protocol or GCP immediately. When an incident report is requested by UCL CTC this should be provided, but an equivalent document (e.g. Trust Incident form) is acceptable where already completed. Where an equivalent document is being submitted to UCL CTC, the patient's trial number must be clearly indicated on all material and any patient identifiers redacted prior to sending, to maintain confidentiality.

If site staff are unsure whether a certain occurrence constitutes a deviation from the protocol or GCP, the UCL CTC trial team can be contacted immediately to discuss.

UCL CTC will use an organisation's history of noncompliance to make decisions on future collaborations.

UCL CTC will assess all incidents to see if they meet the definition of a serious breach.

13.2. Serious Breaches

A "serious breach" is defined as a breach of the protocol or of the conditions or principles of Good Clinical Practice (or equivalent standards for conduct of non CTIMPs) which is likely to affect to a significant degree the safety or physical or mental integrity of the study subjects, or the scientific value of the research.

Systematic or persistent noncompliance by a site with **the principles and conditions of GCP** and/or the protocol, occurring on study within the specified timeframe, may be deemed a serious breach.

In cases where a serious breach has been identified, UCL CTC will inform the REC within 7 calendar days of becoming aware of the breach.

14. STUDY MONITORING AND OVERSIGHT

Participating sites and PIs must agree to allow study related onsite monitoring, Sponsor audits and regulatory inspections by providing direct access to source data/documents as required. Where permitted by site policy, remote access to source data/documents may also be provided by participating sites for remote monitoring by UCL CTC or its representatives.

Patients are informed of this in the patient information sheet and are asked to consent to their medical notes being reviewed by appropriate individuals on the consent form. UCL CTC or its representatives will conduct all monitoring in compliance with the participant consent, site policy and data protection requirements.

UCL CTC will determine the appropriate level and nature of monitoring required based on the objective, purpose, phase, design, size, complexity, endpoints and risks associated with the study. Risk will be assessed on an ongoing basis and adjustments made accordingly.

Details of monitoring activities will be included in the study monitoring plan and conveyed to sites during initiation. The study monitoring plan will be kept under review during the study and updated information provided to sites as necessary.

14.1. Centralised Monitoring

UCL CTC performs centralised monitoring, which requires the submission of documents by sites to UCL CTC for review, including but not limited to: up to date site screening log, current PI GCP and up to date site delegation log. Expectations for document submission will be explained during site initiation and UCL CTC or its representatives will send emails to sites requesting the documents when required.

Sites will be requested to conduct quality control checks of documentation held within the Investigator Site File at the frequency determined for the Study. Checklists detailing the current version/date of version controlled documents will be provided by UCL CTC for this purpose.

14.2. 'Triggered' Onsite/Remote Monitoring

Onsite/remote monitoring visits may be scheduled following UCL CTC review and/or where there is evidence or suspicion of noncompliance at a site with important aspect(s) of the study protocol/GCP requirements.

On site Monitoring

Sites will be sent an email in advance outlining the reason(s) for the visit and confirming when it will take place. The email will include a list of the documents that are to be reviewed, interviews that will be conducted, planned inspections of the facilities and who will be performing the visit.

Remote Monitoring

UCL CTC defines remote monitoring as those monitoring activities conducted at a location remote from the research site which replicate some onsite activities, e.g. source data review. Remote monitoring may be conducted in response to exceptional circumstances preventing access to participating sites (e.g. a global pandemic) or conducted routinely. Details of remote monitoring will be agreed with participating sites, conducted in accordance with site policy and documented in the study monitoring plan.

Sites will be sent an email in advance, confirming when remote monitoring is scheduled to take place and how the source documents will be remotely accessed. The email will include a list of the documents to be reviewed, interviews that will be conducted via telephone/videoconference and who will be performing the remote monitoring.

Remote monitoring will be conducted by UCL CTC or its representatives via a device with adequate security. Patient confidentiality will be maintained at all times, and monitoring activities will be conducted in an appropriate environment where no unauthorised viewing or overhearing of conversations is possible by third parties. Also refer to section 11 (Data Management and Data Handling Guidelines) for details of how source documentation may be submitted to UCL CTC.

Monitoring follow up

Following on site/remote monitoring, the Trial Monitor/Trial Manager will provide a follow up email to the site, which will summarise the documents reviewed and a statement of findings, incidents, deficiencies, conclusions, actions taken and/or actions required. The PI at each site will be responsible for ensuring that monitoring findings are addressed in a timely manner, and by the deadline specified.

14.3. Oversight Committees

14.3.1. Trial Management Group (TMG)

The TMG will include the Chief Investigator, clinicians and experts from relevant specialities and ICONIC study staff from UCL CTC (see page 2). The TMG will be responsible for overseeing the study. The group will meet regularly at least three times a year and will send updates to PIs (via newsletters or at Investigator Meetings).

The TMG will review substantial amendments to the protocol prior to submission to the REC. All PIs will be kept informed of substantial amendments through their nominated responsible individual and are responsible for their prompt implementation.

A TMG charter, which outlines the responsibilities for the ICONIC study, must be signed by all members of the committee before the first meeting is held.

14.3.2. Trial Steering Committee (TSC)

The role of the TSC is to provide overall supervision of the study. The TSC acts on behalf of the funder and the Sponsor.

The ICONIC study will be reviewed by an established UCL CTC TSC that has oversight of a number of trials. All members have signed a TSC charter.

14.3.3. Role of UCL CTC

UCL CTC, on behalf of the sponsor, will be responsible for the day to day coordination and management of the study and the UCL CTC Director will act as custodian of the data generated in the study (on behalf of UCL).

15. WITHDRAWAL OF PATIENTS

In consenting to the study, patients are consenting to assessments, collection of biological samples, follow up and data collection.

15.1. Withdrawal of Consent

If a patient withdraws consent for any aspect of the study, UCL CTC should be notified and the Change of Status eCRF should be completed.

15.1.1. Withdrawal of consent for follow up

If a patient withdraws consent for study follow up but is happy to continue with future data collection from hospital medical notes:

- They will remain on study for follow up
- The patient will no longer have study specific visits and assessments. Follow up eCRFs should be completed based on the routine visit nearest the due date for the follow up eCRF.
- The following eCRFs and data must be submitted at time of withdrawal:
 - Change of Status
 - All eCRFs up to and including the date of withdrawal of consent.

15.1.2. Withdrawal of consent for data collection

If a patient **explicitly** states they do not wish to contribute further data to the study their decision must be respected. The following eCRFs must be submitted at the time of withdrawal of consent:

- Change of Status eCRF
- All eCRFs due up to and including the date withdrawal of consent
- Thereafter, no further data should be submitted.

15.1.3. Withdrawal of consent for use of samples

If a patient withdraws consent for the use of some, or all, of their samples in the study, or for future research, this should be reported on the Change of Status eCRF. Management and data collection should continue as per protocol unless the patient has also withdrawn from study follow up.

15.2. Losses to Follow Up

If a patient moves from the area, every effort should be made for the patient to be followed up at another participating study site and for this new site to take over the responsibility for the patient. Details of participating study sites can be obtained from the UCL CTC trial team, who must be informed of the transfer of care and follow up arrangements.

If it is not possible to transfer to another participating site, the registering site remains responsible for submission of eCRFs.

If a patient is lost to follow up, every effort should be made by the site to contact the patient's GP to obtain information on the patient's status.

At the time of loss to follow up, the following eCRFs should be submitted:

- Change of status eCRF
- All eCRFs due up to and including the date of loss to follow up.

If contact is re-established with the patient, further follow up eCRFs should be submitted, including notifications of relapse and metastases. A death eCRF should also be submitted if the site becomes aware that the patient has died.

Prior to primary analysis and presentation or publication of the primary endpoint data, UCL CTC may ask sites to attempt to re-establish contact with patients who were lost to follow up and/or check hospital records for evidence of when the patient was last known to be alive and evidence of death, disease progression or second malignancies.

15.3. Loss of Capacity

Patients who lose capacity during the study would continue in the study for the purposes of data collection, if appropriate. If the patient regained capacity, an Investigator would discuss with the patient their continued participation in the study and together the patient and Investigator would decide what action, if any, to take.

16. STUDY CLOSURE

16.1. End of Study

For regulatory purposes the end of the study will be defined as two years after recruitment of the last patient, at which point the 'declaration of end of study' form will be submitted to the Ethics Committee, as required and sites notified.

UCL CTC will advise sites on the procedure for closing the study at the site.

Once the end of study has been declared, no more prospective patient data will be collected but sites must cooperate with any data queries regarding existing data to allow for analysis and publication of results.

16.2. Archiving of Study Documentation

At the end of the study, UCL CTC will archive securely all centrally held study related documentation for a minimum of five years. Arrangements for confidential destruction will then be made. It is the responsibility of PIs to ensure data and all essential documents relating to the study held at site are retained securely for a minimum of five years after the end of the study, and in accordance with national legislation.

Essential documents are those which enable both the conduct of the study and the quality of the data produced to be evaluated and show whether the site complied with the principles of GCP and all applicable regulatory requirements.

UCL CTC will notify sites when study documentation held at sites may be archived. All archived documents must continue to be available for inspection by appropriate authorities upon request.

16.3. Early Discontinuation of Study

The study may be stopped before completion as an Urgent Safety Measure on the recommendation of the TSC (see section 14.3.2 Trial Steering Committee (TSC)). Sites will be informed in writing by UCL CTC of reasons for early closure and the actions to be taken with regards the treatment and follow up of patients.

16.4. Withdrawal from Study Participation by a Site

Should a site choose to close to recruitment the PI must inform UCL CTC in writing. Follow up as per protocol must continue for any patients recruited into the study at that site and other responsibilities continue as per the site agreement.

17. STATISTICS

This is a prospective observational cohort study of patients of all ages with histopathologically confirmed OS.

17.1. Sample Size as feasibility endpoint

One of the objectives of Stage 1 was to assess the feasibility of recruiting to this cohort. Our assumption was that 15 sites would be opened during the course of the first 11 months of recruitment. Once all sites were opened (from month 12 onwards), we estimated that the average recruitment rate would be 5 patients/month.

Following the completion of Stage 1 and with the extension to the recruitment period, we expect to recruit a minimum of 300 patients to the study in total.

17.2. Statistical analysis

17.2.1. Stage 1

Descriptive analyses and tabulations have been performed to answer the Stage 1 research questions, as detailed in the table below.

Research question	Analysis
Recruitment objective	
1. Feasibility of recruiting patients across UK paediatric, TYA and adult centres to cohort study	<ul style="list-style-type: none"> Recruitment rate after opening of 15 sites % patients recruited according to age and primary site
Clinical objectives	
2. Determine patterns of treatment of patients across UK treatment sites to inform stage 2 objectives and future studies	<ul style="list-style-type: none"> % patients with MDT documentation of rationale for planned treatment % patients receiving chemotherapy and regimen % patients undergoing surgery % patients receiving radiotherapy and indications
3. Determine feasibility of surgical data capture to inform future studies	<ul style="list-style-type: none"> List of possible reconstruction techniques and their relative frequencies % patients with post-operative complications
4. Imaging: <ul style="list-style-type: none"> determine primary tumour imaging and reporting pre and post chemotherapy Determine variation in imaging modalities used for staging of OS patients to inform future studies 	<ul style="list-style-type: none"> % patients staged with bone scan, PET/CT and/or WB-MRI
Biological objectives	
1. Determine feasibility of fresh frozen tissue sample collection across sites across the UK	<ul style="list-style-type: none"> Collection rate per site Number and % of metastatic/relapse specimens stored for future research % sent for WGS
2. Determine quality of Formalin Fixed paraffin-embedded (FFPE) samples and feasibility of DNA/RNA extraction for validation studies	<ul style="list-style-type: none"> Collection rate of diagnostic/resection FFPE specimens in patients treated with surgery and chemotherapy Audit to determine whether quality and quantity of RNA/DNA sufficient for targeted sequencing and nanostring analysis
3. Evaluate whether two methodologies of circulating tumour cell (CTC) detection can isolate and quantify CTCs in patients undergoing neoadjuvant chemotherapy	<ul style="list-style-type: none"> Identification and quantification of CTC numbers at baseline (pre-treatment) and changes to CTC numbers in response to chemotherapy Descriptive analysis of CTC numbers as histograms and ranges (min, max, IQR, median)

Research question	Analysis
4. Collection of whole blood for potential methylation/ctDNA/future research	<ul style="list-style-type: none"> Collection rate per site
5. Collection of whole blood for Germline DNA analysis	
6. Evaluate collection of serum biomarkers for validation of prognostic scores	<ul style="list-style-type: none"> Collection rate of serum parameters Characterisation of distribution of serum biomarkers as histograms and ranges (min, max, IQR, median)
7. Determine feasibility of using standard pathology reporting to assess margin status for risk of local recurrence	<ul style="list-style-type: none"> Collection rate of pathology reports in patients undergoing surgery Characterisation of tumour size, assessment of resection margins, % necrosis
Patient Reported Outcome	
1. Determine feasibility of data collection for validation of a sarcoma assessment measure (SAM) in patients undergoing treatment for OS	<ul style="list-style-type: none"> Completion rate of SAM, TESS, EORTC-QLQ-C30 and GRC Descriptive analyses of PROs

17.2.2. Stage 2

17.2.2.1: Support and development of ICONIC infrastructure

The first aims for Stage 2 are:

- to support ongoing recruitment to the ICONIC study with additional follow up to form a cohort of a **minimum of 300 patients** with a median of **approximately 56 months follow up**
- to continue the collection of tissues and blood samples for CTC and plasma for circulating DNA (ctDNA)
- to collect peripheral blood mononuclear cells in a cohort of patients for IO research
- to establish an imaging biorepository for collaborative imaging biomarker studies.

Analysis of patient outcomes will be descriptive, using Kaplan-Meier methods for survival outcomes (e.g. overall survival, event-free survival and progression-free survival) and counts/frequencies for categorical outcomes (e.g. pathological response to chemotherapy).

Predictors of patient outcome will be assessed using standard regression techniques: Cox regression for survival outcomes, logistic regression for categorical. We will consider the following potential predictors of patient outcome:

- Blood biomarkers: serum biomarkers and ctDNA/CTCs
- Baseline characteristics (age, performance status, grade, primary site, tumour size, stage)
- Patterns of treatment
- Tissue biomarkers for IO research

Research questions	Planned analysis
Evaluate serum biomarkers (ALP, LDH) as prognostic markers in OS	Quantify serum biomarkers at baseline (pretreatment) and changes in response to chemotherapy. Characterise distribution of serum biomarkers as histograms and ranges (min, max, IQR, median) Assess serum biomarkers for prognostic value by using Cox regression (time-to-event outcomes) and logistic regression (categorical outcomes)
Evaluate ctDNA as predictors of response and outcome in OS	Quantify ctDNA biomarkers at baseline (pretreatment) and changes in response to chemotherapy. Characterise distribution of ctDNA as histograms and ranges (min, max, IQR, median) Assess ctDNA biomarkers for prognostic value by using Cox regression (time-to-event outcomes) and logistic regression (categorical outcomes)
Evaluate whether two methodologies of circulating tumour cell (CTC) detection can isolate and quantify CTCs in patients undergoing neoadjuvant chemotherapy	Identify and quantify CTC numbers at baseline (pretreatment) and changes to CTC numbers in response to chemotherapy Descriptive analysis of CTC numbers as histograms and ranges (min, max, IQR, median)

Power calculations for analysis of biomarkers as predictors of pathological response:

All power calculations assume a fixed 2-sided 5% significance level. As these analyses are hypothesis generating, no adjustments have been made to account for multiple testing.

Stage 1 determined the good pathological response rate to be lower than in clinical trials with 17/43 (37%) having a good response.

Based on data from England (2011-2015), 66% of patients receive surgery and 66% receive chemotherapy. At the outset of Stage 1 we had assumed that 60% of patients receive both treatments. At the end of Stage 1, we determined in our cohort that 72% of patients were planned to receive both modalities of treatment.

We will therefore assume the size of the full dataset of patients treated with both modalities will be at least 65% of 300 patients, thus 195 patients.

1. Serum biomarkers

We expect at least 90% of patients to provide serum samples for biomarker analysis, giving us a cohort size of 176. Assuming an overall pathological response rate of ~40%, and assuming biomarkers will be

categorised based on the median to give an even split (88 per group), below is a summary of the power we will have to detect varying effect sizes:

Alpha	Proportion responding without biomarker (# of responders)	Proportion responding with biomarker (# of responders)	Odds ratio	Risk ratio	Power
0.05	20% (18)	60% (53)	5.9	2.9	99.9%
0.05	25% (22)	55% (48)	3.6	2.2	98.6%
0.05	30% (26)	50% (44)	2.4	1.7	77.8%
0.05	32.5% (29)	47.5% (42)	1.9	1.4	52.9%

2. ctDNA

At the end of Stage 1, 76 patients had provided baseline samples but on treatment samples were fewer and it was not possible for all patients to have baseline samples collected prior to commencing treatment. We therefore assume that with a total sample size of 300, 150 patients will have sufficient pre-samples to correlate biomarkers with clinical characteristics. Assuming an overall pathological response rate of ~40%, and assuming biomarkers will be categorised based on the median to give an even split (75 per group), below is a summary of the power we will have to detect varying effect sizes:

Alpha	Proportion responding without biomarker (# of responders)	Proportion responding with biomarker (# of responders)	Odds ratio	Risk ratio	Power
0.05	20% (15)	60% (45)	6	3	99.9%
0.05	25% (19)	55% (41)	3.6	2.2	97.0%
0.05	30% (23)	50% (38)	2.3	1.7	70.9%
0.05	32.5% (24)	47.5% (36)	2.0	1.5	46.6%

17.2.2.2: Immune oncology

Biomarkers identified through WGS will be summarised using descriptive statistics (median, range, mean, standard deviation) and assessed as prognostic markers for infiltration (poorly infiltrated vs highly infiltrated) using logistic regression. We assume the overall proportion of patients with high infiltration will be 40% and that biomarkers will be categorised based on the median, to ensure equal numbers of those with and without the biomarker. With 2-sided 5% alpha, our power calculations show that with 150 samples we have >70% power to detect a risk ratio (RR) effect size of RR=1.7 or larger and >95% power to detect an effect size of RR=2.2 or larger. We note that the biomarkers in question here are in fact new therapeutic drug targets, where a strong effect size is required to ensure potential therapeutic efficacy, so biomarkers with more marginal effect (e.g. RR~1.1-1.5) are not in scope for the study. Hence, we are confident that with 150 patients in our multiplex immunohistochemistry (mIHC) experiment we are well powered to validate novel targets that are significantly different between poorly and highly infiltrated patient groups.

17.2.2.3: Patient Reported Outcomes

We aim to record patient reported outcomes (PROs) over time which are correlated with clinical data for the purposes of identifying factors including prediagnostic pathways which predict lower quality of life, physical function and impaired emotional wellbeing.

PRO analyses will be largely descriptive, so no power calculation is provided. However, as with all descriptive analyses, the more data we acquire the more likely it is to describe our patient population accurately.

Research questions	Planned analysis
1. Describe longitudinal changes in PRO and functional wellbeing beyond a year after diagnosis	<ul style="list-style-type: none"> Quality of life questionnaires will be analysed using mixed modelling and results reported by each domain with summary statistics. Models will work with the date of assessment, rather than predefined timepoint, to account for varying treatment pathways.
2. Determine the emotional impact of being diagnosed and living with OS	
3. Examine the relationship between prediagnostic intervals and PROs	<ul style="list-style-type: none"> Mixed modelling will be used to assess whether prediagnostic interval is associated with quality of life post diagnosis.
4. Identify factors predicting lower quality of life, physical function and impaired emotional wellbeing.	<ul style="list-style-type: none"> Mixed modelling will be used to assess which factors have an impact on quality of life post diagnosis.

17.3. Transition between Stage 1 and Stage 2

At the end of Stage 1, the analyses described in section 17.2.1 were performed. The analyses proposed for Stage 2 were reviewed during the transition and altered as above based on the findings.

18. ETHICAL CONSIDERATIONS

This study will adhere to the principles and conditions of Good Clinical Practice.

In conducting the study, the Sponsor, UCL CTC and sites shall comply with the protocol and with all relevant guidance, laws and statutes, as amended, applicable to the performance of clinical trials and research including, but not limited to:

- UK Policy Framework for Health and Social Care Research, issued by the Health Research Authority
- Human Rights Act 1998
- Data Protection Act 2018
- General Data Protection Regulation (EU)2016/679 (GDPR)
- Freedom of Information Act 2000
- Human Tissue Act 2004
- Mental Capacity Act 2005

18.1. Ethical Approval

The study will be conducted in accordance with the World Medical Association Declaration of Helsinki entitled 'Ethical Principles for Medical Research Involving Human Subjects' (1996 version) and in accordance with the terms and conditions of the ethical approval given to the study.

The study has received a favourable opinion from the London – Camden and King's Cross Research Ethics Committee (REC) and Health Research Authority (HRA) approval for conduct in the UK.

UCL CTC will submit Annual Progress Reports to the REC, commencing one year from the date of ethical approval for the study.

18.2. Site Approvals

Evidence of assessment of capability and capacity by the Trust or Health Board R&D for a study site must be provided to UCL CTC. Sites will only be activated when all necessary local approvals for the study have been obtained.

18.3. Protocol Amendments

UCL CTC will be responsible for gaining ethical approval for amendments made to the protocol and other study related documents. Once approved, UCL CTC will ensure that all amended documents are distributed to sites as appropriate.

Site staff will be responsible for acknowledging receipt of documents and for implementing all amendments promptly.

18.4. Patient Confidentiality & Data Protection

Patient identifiable data, including initials, age and gender will be collected by UCL CTC. UCL CTC will preserve patient confidentiality and will not disclose or reproduce any information by which patients could be directly identified. Data will be stored in a secure manner and UCL CTC trials are registered in accordance with the Data Protection Act 2018 and GDPR, with the Data Protection Officer at UCL.

19. SPONSORSHIP AND INDEMNITY

19.1. Sponsor Details

Sponsor Name: University College London

Address: Joint Research Office
4th Floor, - West
250 Euston Road
London
NW1 2PG

Contact: Managing Director, UCLH/UCL Research

Tel: 020 3447 9995/2178 (unit admin)
Fax: 020 3447 9937

19.2. Indemnity

University College London holds insurance against claims from participants for injury caused by their participation in the clinical study. Participants may be able to claim compensation if they can prove that UCL has been negligent. However, as this clinical study is being carried out in a hospital, the hospital continues to have a duty of care to the participant of the clinical study. University College London does not accept liability for any breach in the hospital's duty of care, or any negligence on the part of hospital employees. This applies whether the hospital is an NHS Trust or otherwise.

Participants may also be able to claim compensation for injury caused by participation in this clinical study without the need to prove negligence on the part of University College London or another party. Participants who sustain injury and wish to make a claim for compensation should be advised to do so in writing in the first instance to the Chief Investigator, who will pass the claim to the Sponsor's Insurers, via the Sponsor's office.

Hospitals selected to participate in this clinical study shall provide clinical negligence insurance cover for harm caused by their employees and a copy of the relevant insurance policy or summary shall be provided to University College London, upon request.

20. FUNDING

The Bone Cancer Research Trust (BCRT) is supporting the central coordination of the study through UCL CTC.

Research A costs will be reimbursed to sites as per the finance section of the site agreement.

21. PUBLICATION POLICY

All publications and presentations relating to the study should be authorised by the TMG. The TMG will form the basis of the writing committee and advise on the nature of the publications. All collaborators who have actively contributed to the study will be named authors on all main study papers and anyone else who has had a significant input into the conduct, analysis and interpretation of the study.

Specialist papers focusing on a particular aspect of translational research may not require all collaborators to be authors. Data from all sites will be analysed together and published as soon as possible after endpoints have been reached. Participating sites may not publish study results prior to the first publication by the TMG or without prior written consent from the TMG.

The Chief Investigator will make the final decision on authorship. The study data is owned by the Sponsor. The ClinicalTrials.gov number (NCT04132895) should be quoted in any publications resulting from this study.

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APPENDIX 1: REMOTE CONSENT

The following guidance outlining the process for sites to follow if it is not possible for consent to be taken in person. All steps performed need to be documented clearly and filed in patient notes accordingly.

The PIS (as a PDF) and ICF (as a Word document) should be provided to the participant via email or post.

1. There must be an interview between the participant or participant's parent/guardian and the delegated consentor via a phone or videocall

involving a 2 way communication, allowing:

- a. the participant or participant's parent/guardian the opportunity to ask questions and
- b. the investigator to assess their comprehension.

2. The participant or participant's parent/guardian must have ample time to decide whether or not to consent.

So the PIS and ICF should be provided to the participant or participant's parents/guardians **before** the phone or video interview to allow time to read both – although this could be on the same day.

3. The investigator should document in the medical notes that the:

- Participant or participant's parent/guardian has confirmed verbally that they are happy to proceed to consent and that a discussion has been held
- Version numbers of the PIS and ICF used
- Date the PIS and ICF were emailed or posted to the participant
- Date of the patient or participant's parent/guardian signature: this is the date of consent.

4. Confirming the participant's consent:

- The participant or participant's parent/guardian should have the PIS and ICF documents with them for the phone or videocall interview
- The investigator should guide the participant or participant's parent/ guardian through how to agree (or disagree if optional) to each item of the ICF
- The investigator should guide the participant or participant's parent/ guardian through which elements are mandatory for participation and which are optional.

Via email:

- The participant or participant's parent/guardian should initial the applicable boxes, type their name and the date on the consent form and save the document.
- The participant should email the completed ICF back to the investigator.
- The investigator should print the email and the ICF completed by the patient:
 - countersign,
 - date – and:
Add the patient trial number to the ICF once received

Via post:

- The participant or participant's parent/guardian should initial and sign the paper copy of the ICF and send back to the investigator via post.
- The investigator should:
 - countersign,
 - date – and:
Add the patient trial number to the ICF when received.

5. Filing the consent documentation:

The investigator should:

- i. provide a fully completed copy of the ICF to the participant,
- ii. file a copy in the patient medical notes and
- iii. file a copy in the ISF (with participant email as applicable).

APPENDIX 2: QUICK REFERENCE GUIDE TO PATIENT VISITS

SCHEDULE: PATIENTS RECEIVING SURGERY ALONE +/- ADJUVANT CHEMOTHERAPY	Preregistration	Within 28 days after registration	Surgery	End of Treatment (After Adj Chemo, NOT mifamurtide)	Follow up	Relapse (local recurrence or metastases)	For registration	For data collection	For PROs	For Tissue/Blood collection
Histological confirmation of osteosarcoma. Age & sex	X									
Informed consent	X									
Relevant Medical History		X								
Cancer signs & symptoms		X								
Smoking status, Height and Weight		X								
WHO/Lansky PSA		X	X							
Haematology & Biochemistry ^B		X	X	X						
Data collected on primary tumour diagnostic imaging		X								
Data collected on primary tumour biopsy/surgery ^C		X								
Data collected on diagnostic imaging		X								
Osteosarcoma TNM staging ^D		X								
Data collected on treatment plan		X								
Adverse Reactions (in relation to study procedures ONLY) ^E		X		X	X	X				
SAM PRO Booklet (Patients ≥13 only) ^F		X		X	X ^G					
Diagnostic pathway questionnaires ^H		X								
Whole blood sample for germline DNA ^I		X								
Whole blood sample for ctDNA (optional for all patients)		X ^J		X	X ^K	X				
Ship FFPE tumour tissue		X ^L	X ^M			X ^N				
Collect frozen tumour tissue		X ^O	X ^O			X ^O				
Collect information on frozen tumour tissue		X	X	X		X				
Disease status		X	X	X	X					
Pre-operative imaging			X							
Data collected on surgery			X							
Data collected on Adjuvant chemotherapy (if given)				X						
Data collected on radiotherapy including indication (if given)				X						
Survival status					X					
Data collected on relapse (site, treatment etc.) ^P						X ^Q				

- A. WHO PS for patients ≥16 years. Lansky PS for patients <16 years.
- B. Haematology: Hb, ANC, lymphocytes, platelets. Biochemistry: CRP, albumin, ALP, LDH, Creatinine.
- C. If surgery is performed prior to registration.
- D. In accordance with Osteosarcoma TMN version 8, 15/01/18.
- E. For trial specific procedures only e.g. phlebotomy, relapse biopsy if not SOC.
- F. SAM Booklet: Sarcoma Assessment Measure (SAM), EORTC QLQ-C30, Toronto Extremity Salvage Score (TESS) and Global rating of change scale (GRCS) for patients 13 years and over only.
- G. Annually from registration.
- H. Complete within 3 months of registration (by parent/guardian if patient <13).
- I. To be taken at baseline, but can be taken later if this is not possible.
- J. Collect pre surgery.
- K. Collected at clinic visit during follow up no more than 6 monthly.
- L. Diagnostic Archival tissue - collection coordinated by UCL CTC.
- M. From routine surgery taking place after registration - collection coordinated by UCL CTC.
- N. If possible, collected from second sample at relapse if standard of care (SOC), OR after Trial specific consent ONLY If not SOC.
- O. Relapse/Mets Frozen tissue sample will be SOC or optional for ICONIC research - collection coordinated by UCL CTC. Collect at indicated times but send on request
- P. First and any subsequent relapses.
- Q. Continue to collect follow up eCRFs after the first relapse/metastasis form has been completed

SCHEDULE: PATIENTS RECEIVING NEOADJUVANT CHEMOTHERAPY	Preregistration	Baseline: Within 28 days after registration	During Treatment			End of Treatment	Follow up	Relapse (local recurrence or metastases)	<div style="display: flex; flex-direction: column; gap: 5px;"> <div style="background-color: #ffffcc; border: 1px solid black; width: 20px; height: 10px; margin-bottom: 5px;"></div> For registration <div style="background-color: #ccffcc; border: 1px solid black; width: 20px; height: 10px; margin-bottom: 5px;"></div> For data collection <div style="background-color: #ccccff; border: 1px solid black; width: 20px; height: 10px; margin-bottom: 5px;"></div> For PROs <div style="background-color: #ffcccc; border: 1px solid black; width: 20px; height: 10px;"></div> For Tissue/Blood collection </div>
			End of Neoadjuvant chemotherapy	Surgery	End of Adjuvant chemotherapy (NOT mifamurtide)				
Histological confirmation of osteosarcoma. Age & sex	X								<p>A. WHO PS for patients ≥16 years. Lansky PS for patients <16 years.</p> <p>B. Haematology: Hb, ANC, lymphocytes, platelets. Biochemistry: CRP, albumin, ALP, LDH, Creatinine.</p> <p>C. If surgery is performed prior to registration.</p> <p>D. In accordance with Osteosarcoma TMN version 8, 15/01/18.</p> <p>E. For trial specific procedures only e.g. phlebotomy, relapse biopsy if not SOC.</p> <p>F. SAM Booklet: Sarcoma Assessment Measure (SAM), EORTC QLQ-C30, Toronto Extremity Salvage Score (TESS) and Global rating of change scale (GRCS) for patients 13 years and over only.</p> <p>G. Annually from registration.</p> <p>H. To be completed within 3 months of registration, by parent/guardian if patient <13.</p> <p>I. To be taken at baseline, but can be taken later if this is not possible.</p> <p>J. For at least 50 neoadjuvant chemotherapy patients.</p> <p>K. If MAP chemo: collect 7-0 days prior to C2D21 methotrexate; If AP or other chemo: collect pre Cycle 3 (ideally up to 7 days prior/14 days post Cycle 2)</p> <p>L. If not taken at end of neoadjuvant chemotherapy.</p> <p>M. Please take afterwards if not possible in 28 days after registration/before treatment.</p> <p>N. Before surgery if possible, if not taken at end of neoadjuvant chemotherapy.</p> <p>O. Collected at clinic visit during follow up no more than 6 monthly.</p> <p>P. Diagnostic Archival: collection coordinated by UCL CTC, shipped by surgery site or traced/shipped from Chemo site as soon as possible after registration, or on request.</p> <p>Q. From routine surgery taking place after registration, collected from surgery sites.</p> <p>R. If possible, collected from second sample at relapse if standard of care (SOC), OR after Trial specific consent ONLY If not SOC.</p> <p>S. Relapse/Mets Frozen tissue sample will be SOC or Optional for ICONIC research - collection coordinated by UCL CTC. Collect at indicated times but send on request.</p> <p>T. First and any subsequent relapses.</p> <p>U. Continue to collect follow up eCRFs after the first relapse/metastasis form has been completed.</p>
Informed consent	X								
Relevant Medical History		X							
Cancer signs & symptoms		X							
Smoking status, Height and Weight		X							
WHO/Lansky PSA ^A		X		X					
Haematology & Biochemistry ^B		X		X	X				
Data collected on primary tumour diagnostic imaging		X							
Data collected on primary tumour biopsy/surgery ^C		X							
Data collected on diagnostic imaging		X							
Osteosarcoma TNM staging ^D		X							
Data collected on treatment plan		X							
Adverse Reactions (in relation to study procedures ONLY) ^E		X	X	X	X	X	X		
SAM PRO booklet (Patients ≥13 only) ^F		X			X	X ^G			
Diagnostic pathway questionnaires ^H		X							
Blood sample for germline DNA ^I		X							
Blood sample for CTCs ^J		X	X ^K	X ^L			X		
Whole blood ctDNA sample (optional for all patients)		X ^M	X	X ^N	X	X ^O	X		
PBMC (for 20 UCLH patients only)		X	X	X ^N	X		X		
Ship FFPE tumour tissue		X ^P		X ^Q			X ^R		
Collect frozen tumour tissue		X ^S		X ^S			X ^S		
Collect information on frozen tumour tissue		X		X	X		X		
Data collected on chemo regimen (dose & dates)			X		X				
Disease status (if performed)			X	X	X	X			
Pre-operative imaging				X					
Data collected on surgery				X					
Data collected on chemotherapy given			X	X					
Data collected on radiotherapy including indication (if given)					X				
Survival status						X			
Data collected on relapse (site, treatment etc.) ^T							X ^U		

APPENDIX 3: PERFORMANCE STATUS

WHO Performance Status

Grade	Description
0	Able to carry out all normal activity without restriction
1	Restricted in physically strenuous activity but ambulatory and able to carry out light work
2	Ambulatory and capable of all self-care but unable to carry out any work; up and about more than 50% of waking hours
3	Capable of only limited self-care; confined to bed or chair more than 50% of waking hours
4	Completely disabled; cannot carry out any self-care; totally confined to bed or chair

Lansky Performance Status

	Lansky score (Age: <16 years)
100	Fully active, normal.
90	Minor restrictions in physically strenuous activity.
80	Active, but tires more quickly.
70	Both greater restriction of, and less time spent in, active play.
60	Up and around, but minimal active play; keeps busy with quieter activities.
50	Gets dressed, but lies around much of the day; no active play; able to participate in all quiet play and activities.
40	Mostly in bed; participates in quiet activities.
30	In bed; needs assistance even for quiet play.
20	Often sleeping; play entirely limited to very passive activity.
10	No play does not get out of bed. Moribund.
0	Unresponsive. Dead.

APPENDIX 4: STAGE 1 OBJECTIVES AND ANALYSIS

Objectives	Endpoint Analyses
Recruitment	
1. Recruit patients across UK paediatric, TYA and adult centres to cohort study <ul style="list-style-type: none"> ➤ Open at least 15 sites representative across all age groups across UK ➤ Recruitment rate: 5 patients/month when 15 sites open 	<ul style="list-style-type: none"> • Calculation of recruitment rate after opening of 15 sites • Tabulation of % patients recruited according to age and primary site
Clinical data collection	
1. Determine patterns of treatment of patients across UK treatment sites <ul style="list-style-type: none"> ➤ Establish number and % of patients receiving systemic therapy across age cohorts ➤ Establish number and % patients undergoing surgery, ➤ Establish list of reasons for inoperability and their relative frequencies ➤ Establish current indications for RT and their relative frequencies 	Tabulations of: <ul style="list-style-type: none"> • % patients with MDT documentation of rationale for planned treatment • % patients receiving chemotherapy and regimen • % patients undergoing surgery • % patients receiving radiotherapy and indications
2. Determine feasibility of surgical data capture on impact of surgery on complication rate and function <ul style="list-style-type: none"> ➤ Establish variation in reconstruction techniques ➤ Determine number and % patients with post-operative complications 	Tabulations of: <ul style="list-style-type: none"> • possible reconstruction techniques and relative frequencies • % patients with post-operative complications
3. Imaging: Determine feasibility of establishment of an imaging repository to <ul style="list-style-type: none"> ➤ Evaluate conformance of primary tumour imaging and reporting of response to chemotherapy ➤ Evaluate variation in imaging modalities utilised for staging of OS patients 	Tabulation <ul style="list-style-type: none"> • % of scans available for central radiology review • % of reports available for review • % patients staged with bone scan, PET/CT and/or WB-MRI
Biological data collection	
1. Sample collection of fresh frozen tissue across UK sites <ul style="list-style-type: none"> ➤ Collect diagnostic/resection specimens in 50% patients recruited 	Calculation of <ul style="list-style-type: none"> • collection rate per site • number and % of metastatic/relapse specimens stored for future research • % of specimens sent for WGS

Objectives	Endpoint Analyses
2. Determine quality of Formalin Fixed paraffin-embedded (FFPE) samples and feasibility of DNA/RNA extraction for validation studies ➤ Collect 80% of diagnostic/resection FFPE specimens in patients receiving chemotherapy and surgery	Calculation of collection rate of diagnostic/resection FFPE specimens in patients treated with surgery and chemotherapy • Audit to determine whether quality and quantity of RNA/DNA is sufficient for targeted sequencing and nanostring analysis
3. Evaluate whether the two methodologies of circulating tumour cell (CTC) detection can isolate and quantify CTCs in patients undergoing neoadjuvant chemotherapy Collect pre and post neoadjuvant chemotherapy blood samples in 50 patients to evaluate isolation and quantification of CTC using DEPAArray™ and flow cytometry	• Identification and quantification of CTC numbers at baseline (pretreatment) and after chemotherapy • Descriptive analysis of CTC numbers as histograms and ranges (min, max, IQR, median)
4. Collection of plasma for potential methylation/ctDNA/future research	• Calculation of collection rate per site
5. Evaluate collection of serum biomarkers for validation of prognostic scores ➤ Collect serum parameters in 90% patients	• Calculation of collection rate of serum parameters • Characterisation of distribution of serum biomarkers as histograms and ranges (min, max, IQR, median)
6. Determine feasibility of using standard pathology reporting to determine risk of local recurrence ➤ Collect pathology reports on 90% patients undergoing surgery	• Characterisation of tumour size, measurement of narrowest surgical margin status, pathological response to chemotherapy (% necrosis)
PROMs data collection	
1. Collect data for validation of a sarcoma assessment measure (SAM) in patients undergoing treatment for OS ➤ Completion of SAM, TESS, EORTC QLQ-C30 and Global rating of change (GRC) in 60% of patients > 13 years at 2 time points	• Calculation of completion rate of SAM, TESS, EORTC QLQ-C30 and GRC. • Descriptive analyses of PROMs at both time points
Osteosarcoma and routes to diagnosis	
1. Determine feasibility of patients and GPs returning questionnaires ➤ All patients recruited given questionnaire GPs sent questionnaire	• % of patients returning questionnaires • % of GPs returning questionnaires

APPENDIX 5: PROTOCOL VERSION HISTORY

Protocol:		Amendments:		
Version no.	Date	Amendment no.	Protocol Section (no./title)	Summary of main changes from previous version.
1	07/03/2019	n/a	n/a	n/a
2	23/01/2020	2	Throughout	References to <i>snap frozen</i> changed to <i>fresh frozen according to local policy</i> . Administrative changes - general updates, minor clarifications, correction of typos/formatting errors
			1.1, 6.2.2	Change exclusion criteria from: • Diagnosis more than <i>three</i> months prior to registration To: • Diagnosis more than <i>four</i> months prior to registration
			9.2.3	Request that the early diagnosis questionnaire should be completed within 3 months of diagnosis changed to within 3 months of registration. Parent or guardian should complete the Early Diagnosis Questionnaire for patients under the age of 13.
			9.2.4	Added: NB if patient has started chemotherapy prior to study entry this baseline sample may still be collected)
			10.1.4	First paragraph changed from: All FFPE samples will be stored at RNOH. Sites may request return of routine samples if required for clinical purposes, but would be requested to return the samples to RNOH. To: All FFPE samples will be stored at RNOH. <i>During the Study</i> sites may request return of routine samples if required for clinical purposes but would be requested to return the samples to RNOH after this. <i>At the end of the Study all archival FFPE samples will be returned to site.</i>

Protocol:		Amendments:		
Version no.	Date	Amendment no.	Protocol Section (no./title)	Summary of main changes from previous version.
			10.3	Changed to: A new biopsy, FFPE and snap fresh frozen according to local policy if possible, should be obtained at relapse/resection of local recurrence or metastases for the study if <i>not</i> done as part of standard of care. This is optional for patient.
3	25 June 2021	5	Appendix 1:	Was Abbreviations, this now moved to start of protocol.
			Appendix 2:	Now Remote consent: added following Covid Lockdowns.
				Quick reference guide to patient visits: amended to include changes to Sections 9 and 10 in version 2, 23 January 2020
			Trial summary, section 9	SAM booklet to be completed at registration
			5.2	Additional information for patients lacking capacity to consent added.
			5.3	Remote consent added
			9 and 10	Clarification that whole blood samples are taken at site Removal of 'at least 50 patients' for CTC sample collection. Confirmation that bloods should be taken at clinic follow up, no more than 6 monthly Clarification for procedure for submission of CTC blood samples Clarification that ctDNA samples are to be taken for all consenting patients
	14	Instructions on remote monitoring updated		
	15	Clarification on withdrawal of patients		

Protocol:		Amendments:		
Version no.	Date	Amendment no.	Protocol Section (no./title)	Summary of main changes from previous version.
4	23 Sept 2022	7	1.1	<p>Stage 2 main objectives: 'To enrol at least 160 patients' changed to 'to enrol 160 – 220 patients' Target accrual stage 2: Change of total number of patients from ' minimum 160 patients recruited over 3 years' to '160 – 220 patients recruited over 3 years 3 months' Duration of recruitment: Changed from '3 years' to '3 years 3 months'</p>
			17.1	<p>'Since recruitment is expected to last for 3 years in total (Stage 1 + Stage 2), the total expected sample size will be at least 160 patients' changed to: 'Since recruitment is expected to last for 3 years 3 months in total (Stage 1 + Stage 2), the total expected sample size will be 160 - 220 patients'</p>
5	17 March 2023	10		Administrative changes - general updates, minor clarifications, correction of typos/formatting errors
			TMG	<p>Removal of Professor Jeremy Whelan, Dr Asif Saifuddin, Laura White, Andre Lopes and Dr Phillip Green from the list of members of the TMG. Addition of Mr Michael Parry, Mr Jonathan Stevenson, Dr Jenny Sherriff, Dr Kevin Litchfield, Dr Lorna Fern, William Wilson and Dr Filipa Vance to the list of members of the TMG.</p>

Protocol:		Amendments:		
Version no.	Date	Amendment no.	Protocol Section (no./title)	Summary of main changes from previous version.
			1.1	Addition of <ul style="list-style-type: none"> • Can Germline DNA alterations be confirmed in OS? • Can WGS samples be collected for the OS population? To stage 2 objectives <ul style="list-style-type: none"> • Change of end of study definition to 2 years after recruitment of last patient. • Change from To enrol 160 – 220 patients to To enrol 300-350 patients. (and at all other sections of protocol referring to recruitment targets) To duration of recruitment <ul style="list-style-type: none"> • From 3 years 3 months to 31 January 2025. To duration of follow up To minimum of two years, with a median of approximately 56 months follow up.
			1.2	Addition of study flowchart. Addition of Sample Schedules tables.
			2.2.4	Addition of section on Immune Oncology research.
			3.1.1	Deletion of Stage 1 objectives table which has become appendix 4. Addition of collection of section 5: whole blood for germline DNA analysis and Section 6: Rate of WGS samples analysis across the cohort to biological data collection objectives
			3.1.2	Addition of new Stage 2 objectives
			4.1.2	Deletion of requirement for GCP training as ICONIC is a non-CTIMP observational study.
			5.2	Addition of This Clause is not to be applied at any Scottish or Northern Irish ICONIC site. To start of section.
			6.2	Addition of Patients who do not consent to completion of PROs but are willing to donate blood and/or tissue samples are eligible for the Study.
			9	Addition of collection of PMBC samples for 20 UCLH patients throughout section 9

Protocol:		Amendments:		
Version no.	Date	Amendment no.	Protocol Section (no./title)	Summary of main changes from previous version.
			9.2.2	Addition of The SAM booklet (Sarcoma Assessment Measure (SAM), EORTC-QLQ-C30, TESS and GRC) can be completed over the phone as well as by post or in person. To end of first paragraph of section.
			9.2.3	Addition of It must be completed in person and not undertaken over the phone or posted To first paragraph of section.
			9.2.4	Addition of • If the baseline ctDNA sample is missed for any reason, please continue with the on treatment, end of treatment and follow up samples. To list of conditions for all patients.
			9.3.2	Addition of • If the baseline ctDNA sample was missed for any reason, please continue with the on treatment, end of treatment and follow up samples To list of conditions for all patients
			9.6.2	Addition of • If the baseline optional ctDNA sample was missed for any reason, please continue with the on treatment, end of treatment and follow up samples To list of conditions for all patients.
			10.2	Addition of We anticipate collection of fresh frozen tumour specimens, where possible, on request. At the end of the Study all routine fresh frozen tumour specimens will be returned to site. After first paragraph.
			10.7	Addition of section on collection of Whole blood samples for PBMCs
			14.2	Addition of section on Remote monitoring.
			14.3.3	The UCL CTC Director is named as custodian of data generated in the study.

Protocol:		Amendments:		
Version no.	Date	Amendment no.	Protocol Section (no./title)	Summary of main changes from previous version.
			16.1	Amendment of definition of end of study from 4 years after start of recruitment to 2 years after recruitment of last patient.
			17.2.1	Addition of 5. collection of plasma for Germline DNA analysis To research question section of table.
			17.2.2	Addition of updated Stage 2 outcomes to 17.2: Statistical analysis.
			19.1	Revision of address of Sponsor and indemnity details.
			Appendix 2	Addition of <ul style="list-style-type: none"> • Q. continue to collect follow up after the first relapse/metastasis form has been completed And <ul style="list-style-type: none"> • U. continue to collect follow up after the first relapse/metastasis form has been completed. To each table of appendix. <ul style="list-style-type: none"> • PBMC collection to table 2 of appendix.
			Appendix 4	Stage 1 objectives and analysis moved from section 3.1.1. Change from plasma to whole blood in section 4 of table.
6		14		Administrative changes - general updates, minor clarifications, correction of typos/formatting errors
			TMG	Removal of Dr Kevin Litchfield, Sharon Forsyth and Kate Sparksman from the list of members of the TMG. Addition of Temi Adedoyin, Rubina Begum, Alan Sahin and Molly Richardson to the list of members of the TMG.
				Addition throughout of <ul style="list-style-type: none"> • New instructions for sending ctDNA samples to ECMC GCLP CI Lab.
			Section 10	Addition of Samples flowchart